

# Sterilization depyrogenation



*Institute of Pharmaceutical Technology and Biopharmacy*

# Sterilization

Sterilization is an **operation** that can **remove/destroy** the micro-organisms **from/in** the product.

**Aim:** sterility of the preparation.

**Sterility (sterile prod.):** „is the absence of viable micro-organisms.“  
(viable- and spore-forms)

**Sterility control:**

- proper, validated procedure (GMP)
- examination
- unopened product / container

# Sterilization

**Sterilization:** the summation of operations and procedures, that can ensure sterility of the final-product.

„The chosen test **method depends on the properties of the material to be sterilized.**

Eventual heat-sensitivity must be taken into consideration, and sterility must be achieved without causing change in the therapeutic action.”

„**The sterility of a product cannot be guaranteed by testing; it has to be assured by the application of a suitably validated production process.**”

# Sterilization

## When a fully validated thermal sterilization method:

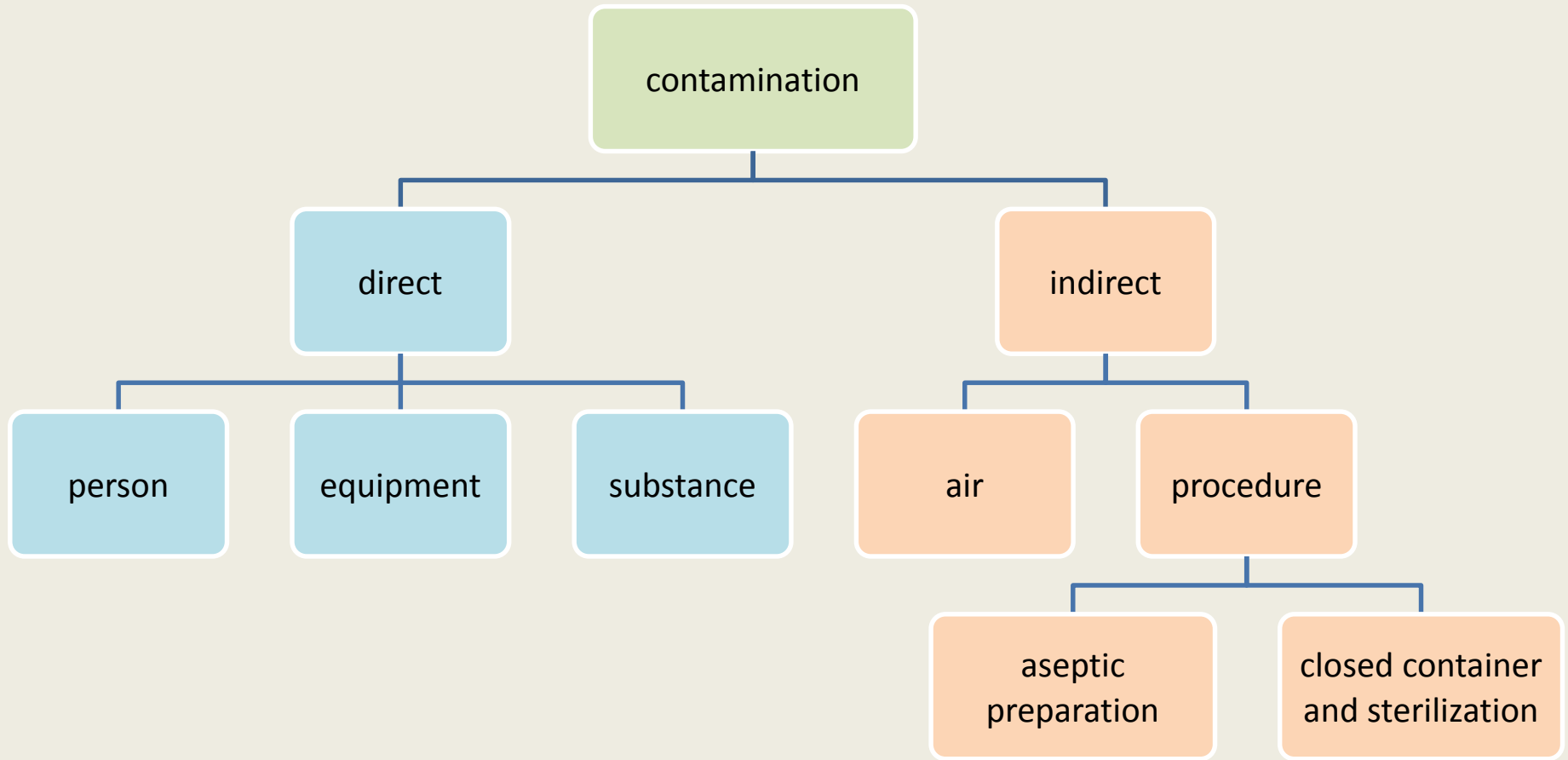
by steam, dry heat or ionising radiation is used, **parametric release**, that is a release of a batch of sterilized items **based on process data rather than on the basis of submitting a sample of the items to sterility testing**, may be carried out, subject to the approval of the competent authority.

## OR:

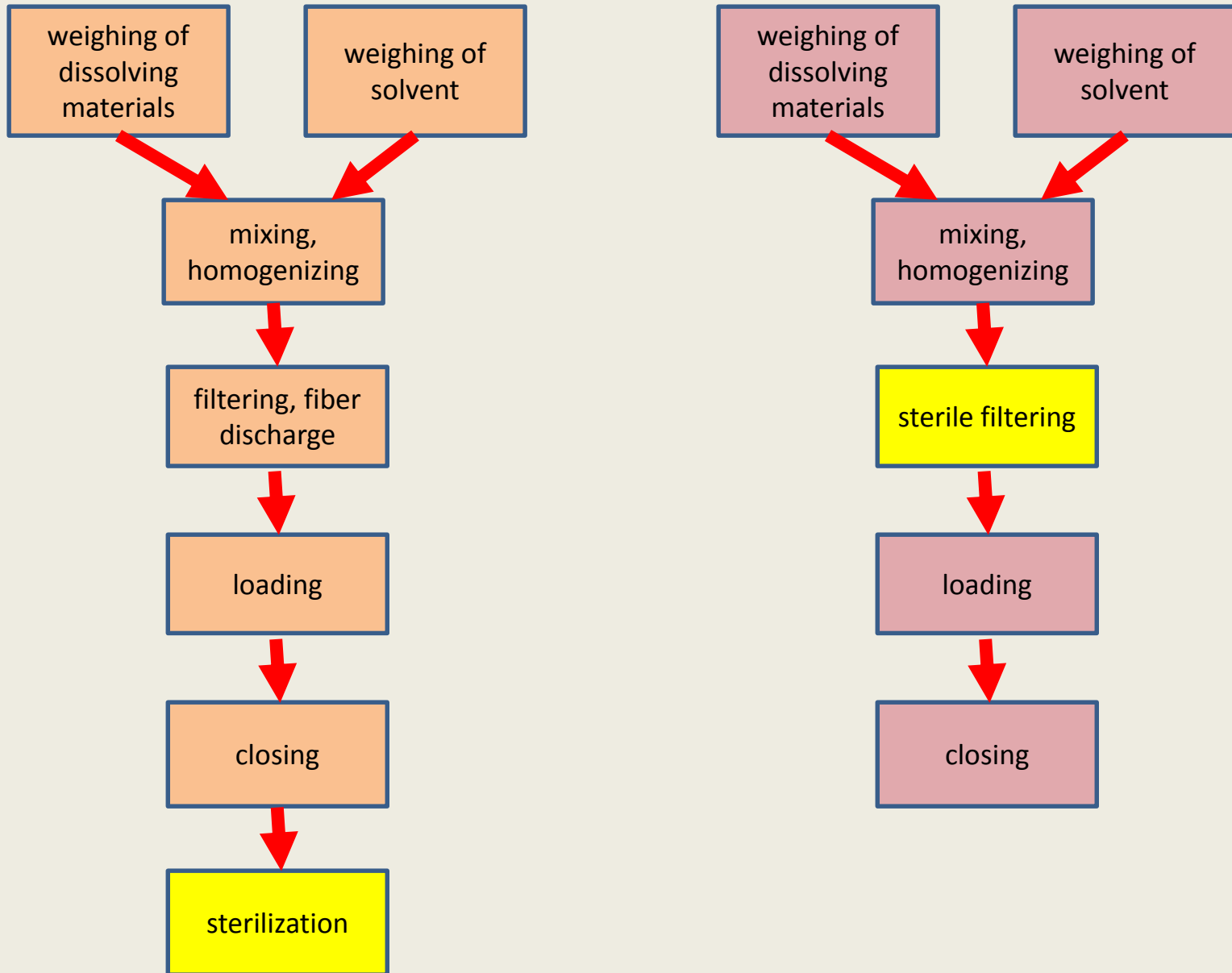
- „If **thermal sterilisation is not possible, filtration through a bacteria-retentive filter or aseptic processing is used**; wherever possible, appropriate additional treatment of the product ... in its final container is applied.”

„In all cases, the container and closure are required to maintain the sterility of product throughout its self-time.”

# Contamination's options



# Procedures



# Principles of sterilization

## Micro-organisms

- **Elimination:**        **filtration**
- **Damage:**            physical methods: **heat, radiation**  
                              chemical methods: **oxydation**

## Effectivness:

- Resistance of micro-organisms
- Number of bacteria
- Effectivness of the method

$$K = \frac{1}{t} \ln \frac{B}{b}$$

K = damage rate

t = sterilization time

B = numbers of bacteria at the beginning

b = numbers of bacteria at the t-time

# Background of sterilization

Ph.Eur.6: „**SAL**” (**S**terility **A**ssurance **L**evel)

„The SAL of sterilizing process is the degree of assurance with which the process in question renders a population of items sterile.”

„The SAL in a given process is expressed as the probability of non-sterile item in that population.”

*„An SAL of  $10^{-6}$ , for example, denotes a probability of not more than one viable micro-organism in  $1 \times 10^6$  sterilized items of the final product.”*

The inactivation of micro-organisms follows an exponential statistic law.

$$K = \frac{1}{t} \ln \frac{B}{b}$$

K = damage rate

t = sterilization time

B = numbers of bacteria at the beginning

b = numbers of bacteria at the t-time



# Definition of „Sterile”

- A sterilization process must deliver a Sterility Assurance Level (SAL) of 1 in a million ( $10^{-6}$ )
- It is not possible to measure  $10^{-6}$
- The required SAL can be achieved by **applying a process that will reduce the number of organisms to zero** and then apply a safety factor that will deliver an extra 6 log reduction

# Background of sterilization

„The inactivation of micro-organisms by physical or chemical means follows an exponential law; thus there is always a finite statistical probability that a micro-organism may survive the sterilizing process.” .

$$\text{Higuchi-Buss} \quad \rightarrow \quad \ln t_d = \Delta H_s / R T + K$$

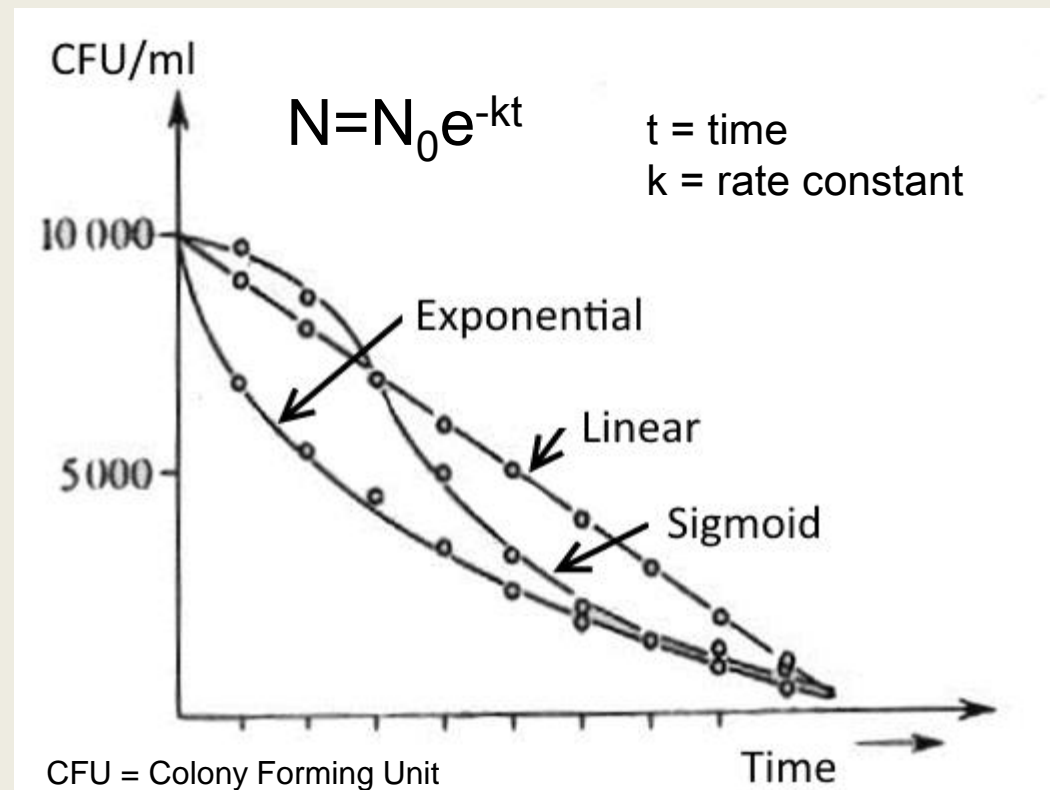
$\Delta H_s$  = necessary heat to sterilization in present one determined species

$K$  = species constant

$T_d$  = sterilization time

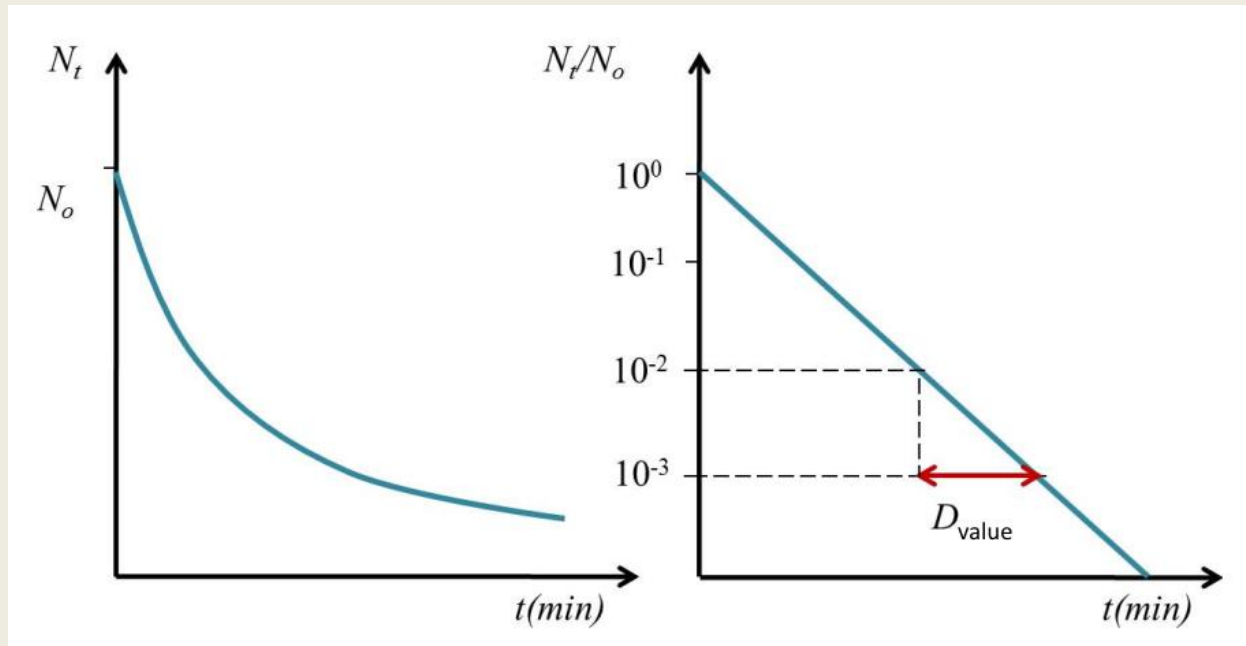


- numbers of micro-organisms (N),
- resistance,
- enviroment,
- Lower concentration of salts
- pH~7,
- Lower temperature



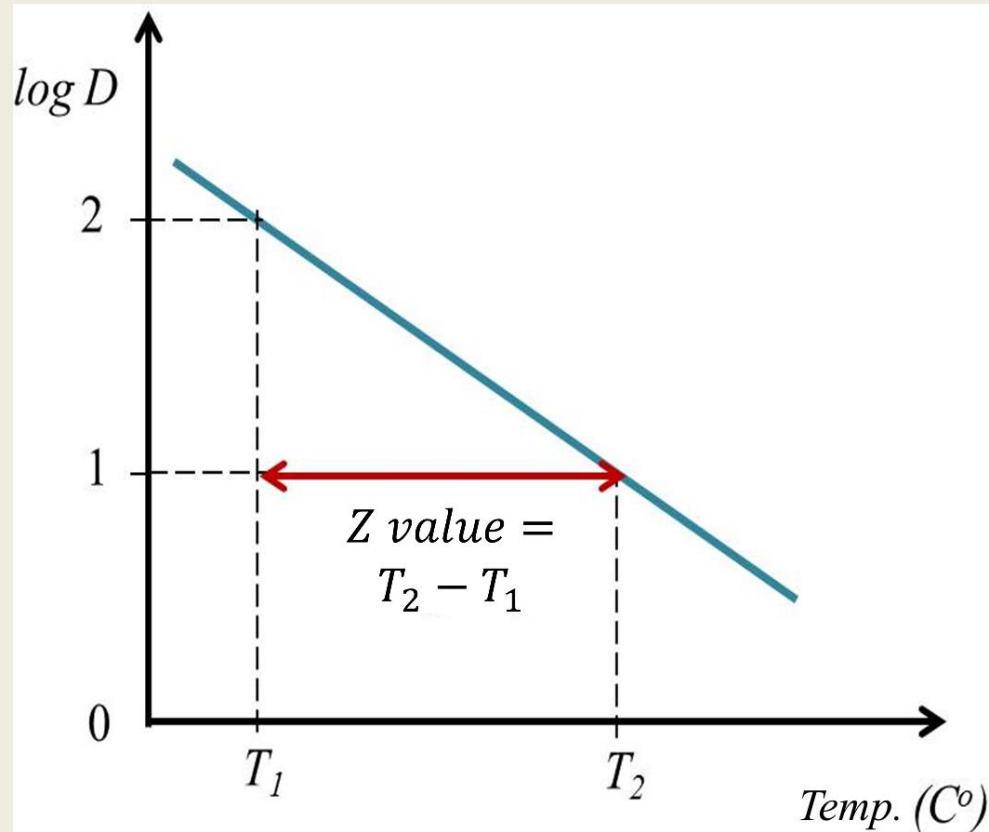
# Definition of „Sterile”

Resistance of an organism is referred as its “**D-value**”



**D-value** - Time (or dose) **required to reduce the population of organisms by 1/10 of the initial number of microbes** (or 90%).

# Definition of „Sterile”



The rate of heat death of microorganisms varies with temperature.  
**The Z value is the temperature change that changes the D value by an order of magnitude.**

# Methods of sterilization

## I. Heat-sterilization

(flame sterilization (Ph. Hg. VI))

dry heat sterilization

steam sterilization

## II. Radiation

IR, UV, ionisation

## III. Filtration

adsorption and sieve effect

## IV. Chemical routes

formaldehyde,  $\beta$ -propiolactone, ethylen-oxyde,

## V. Plasm sterilization

hydrogen-peroxide plasm

# Sterilization procedure

- Registration of the critical parameters (pressure and temperature) is important
- „The **location within the sterilizing chamber that is least accessible to the sterilizing agent is determined** for each loading configuration of each type and size of container or package *(for example, the coolest location in an autoclave).*”
- It is advisable to **take the sensors to the coolest parts** of the chamber.
- Sensors can be filled into bottles, too.

# Dry heat sterilization

(Ph.Hg.VII.)

Dry heat sterilization		
°C	Time	
	Ventillation (+/-)	
	+	-
200 ±5	10	35
180 ±5	25	60
160 ±5	45	120
140±5	-	180

Ph.Eur.6:

min. 160°C , min 2h

Proper reproductivity level

→ SAL 10<sup>-6</sup> or better

- Temperature-sensing elements on the coolest part of loaded sterilizer.
- Biological indicators

above 220°C → sterilization + depyrogenation

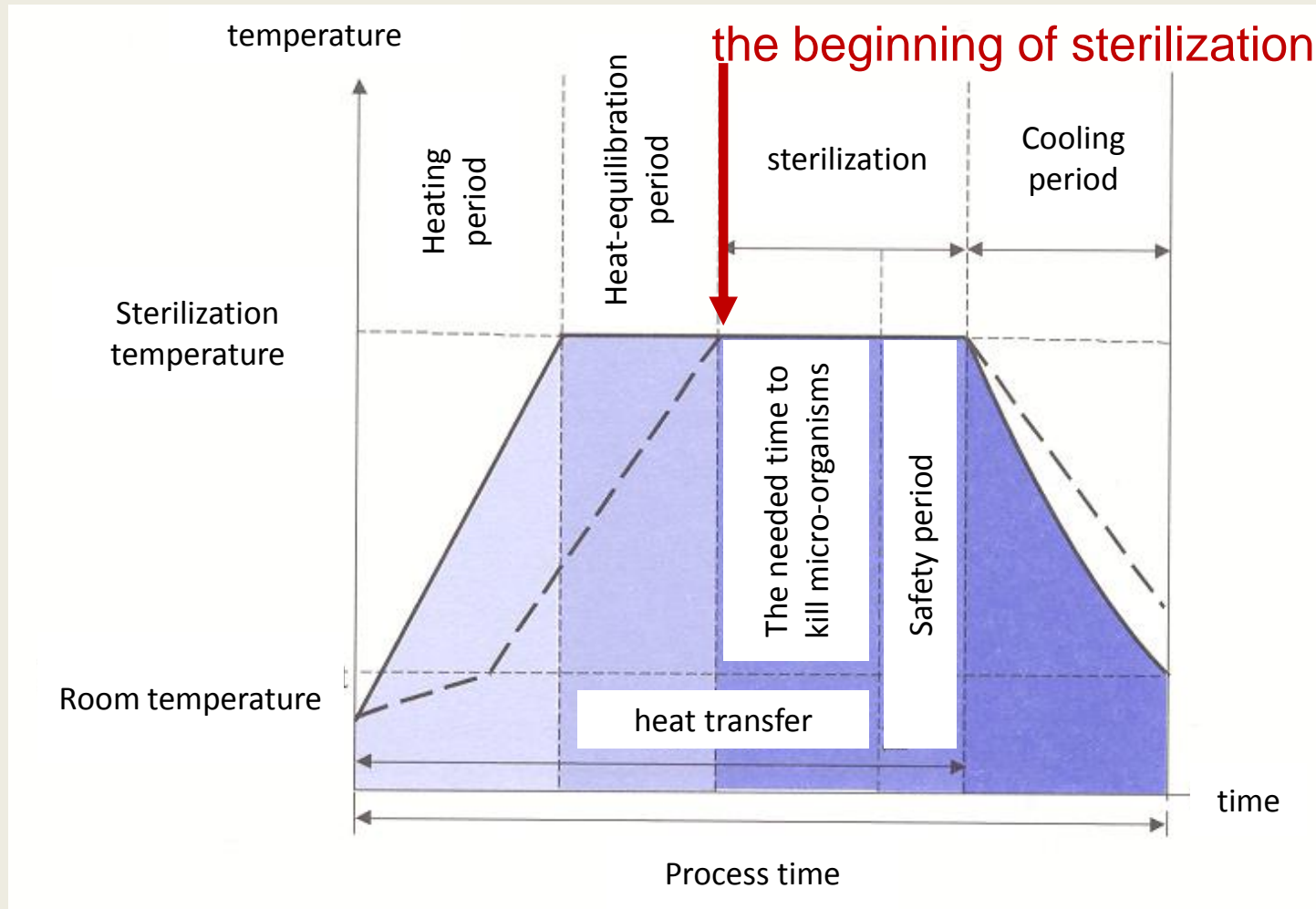
Registration of pressure and temperature

Application of microbial indicators

Above 220°C sterilization of glasswares and depyrogenation

# Dry heat sterilization

## Procedure of dry-heat sterilization





# Steam sterilization (heating in an autoclave)

Ph.Hg.VII

Autoclave		
°C	atm	min
134 ±2	+2.1	10
121 ±3	+1.1	20

Ph.Eur. 6:

Under pressure.

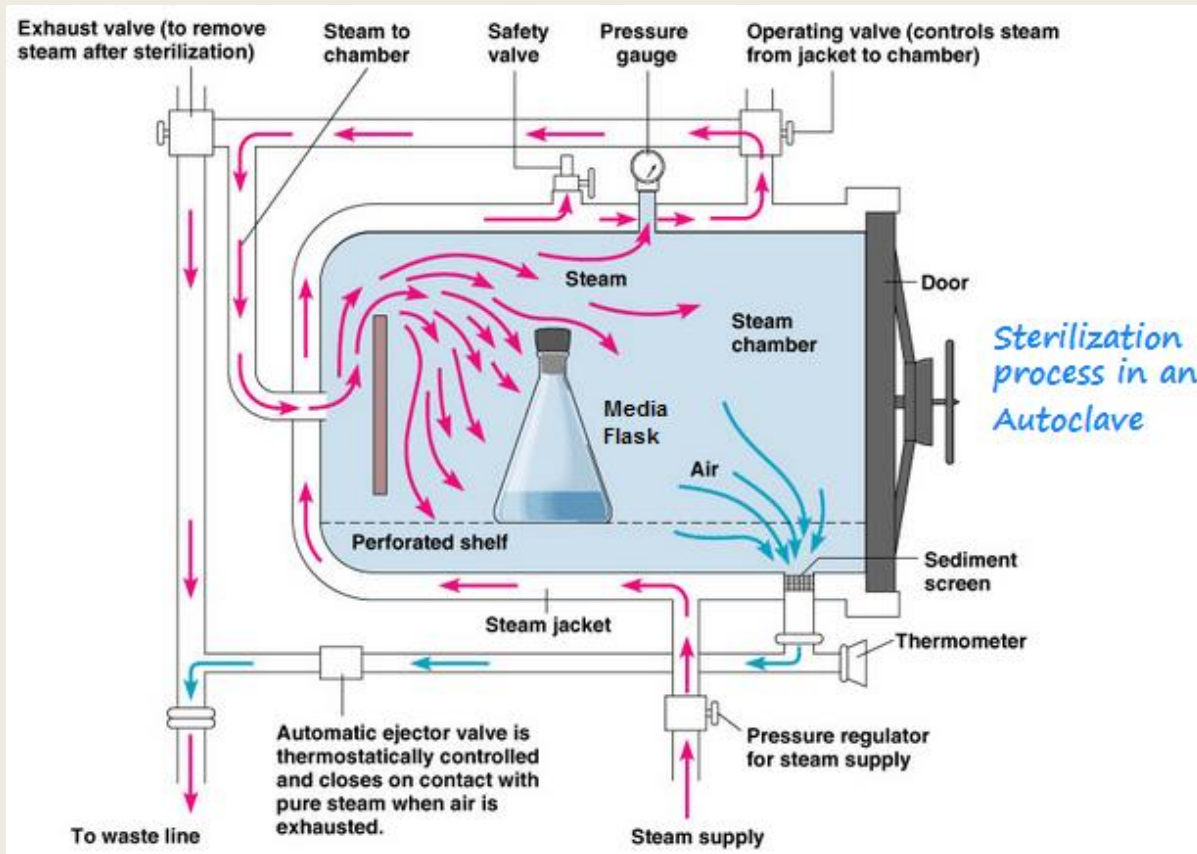
Applicable, especially for aqueous preparations.(min: 15min,  $\geq 125^{\circ}\text{C}$ )

SAL  $10^{-6}$  or better

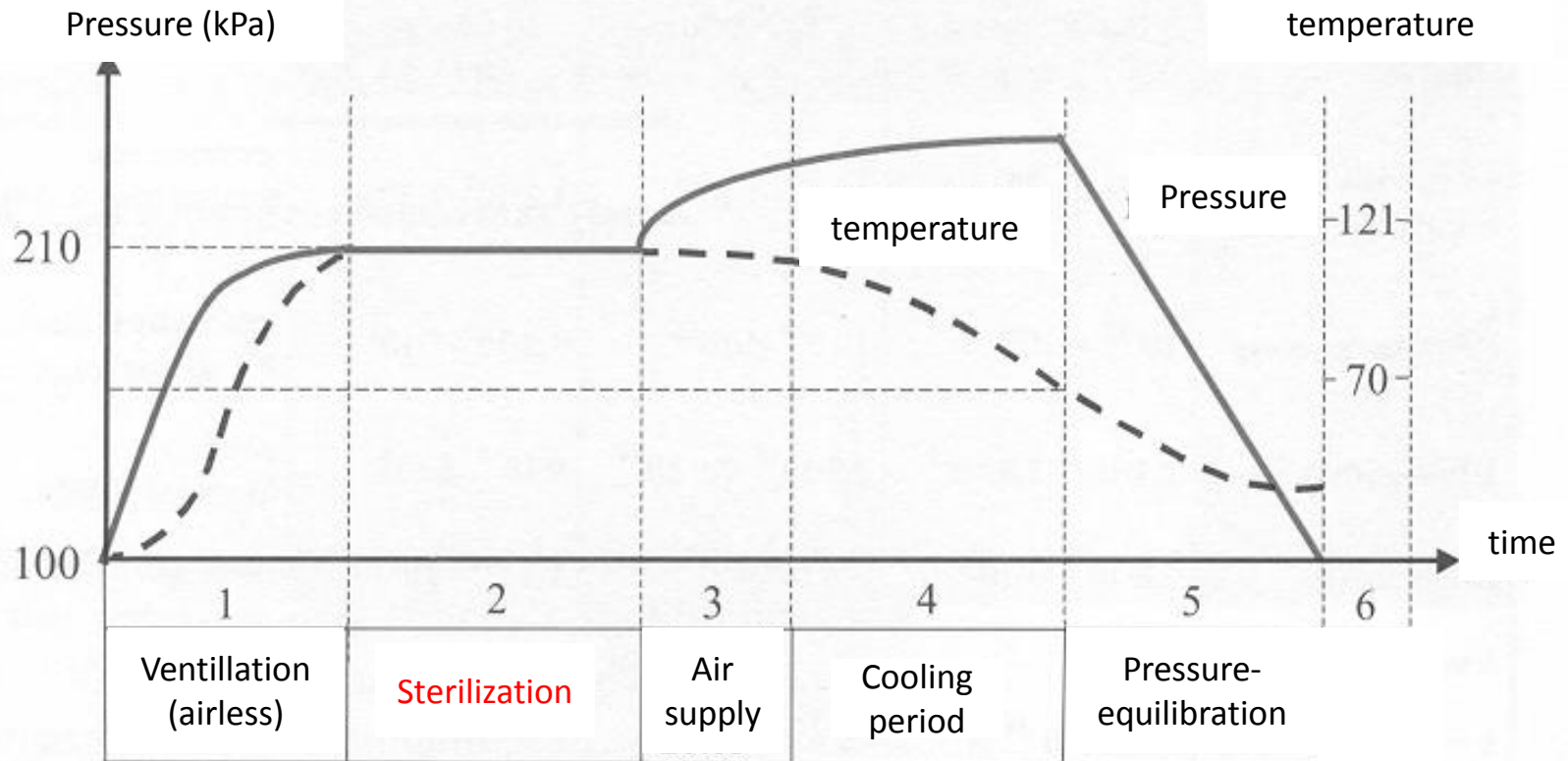
Registration of temperature and pressure

Microbial indicators

# Steam sterilization (heating in an autoclave)



# Steam sterilization (heating in an autoclave)



# Radiation

## Non-ionising radiation

### Ultraviolet (UV)

- 200-400nm
- The most effective range: 240-280nm (germicide:253,7 nm)
- Aseptic work area, and processing
- Manipulator, lamina(i)r box
- Disadvantages: the glass absorbs the radiated beam (surface effect)

### Infrared (IR)

- heat effect

## UV-radiation

Applied by aseptic work places. (germicide)

## IR-radiation

The temperature of the system is increased.  
The heat-sensitive microorganisms will be damaged .

## Ionising radiation: X-ray, $\alpha$ -, $\beta$ -, $\gamma$ -radiation

- Ph.Hg.VIII. (terminal sterilization)
- source: Co 60 or electrone-beam sterilizer

Water based solution

Oxidizing radicals

Irreversible changes in protein's structure

Molecular damages (the active ingredient too)

# Radiation

## Ionising radiation sterilization (Ph.Hg.VIII.:)

- ionising radiation
- radioisotropic source (cobalt 60) – gamma-  
or by electron accelerator
- standard: absorbed dose : 25 kGy (kgray= $\text{J kg}^{-1}$ )
- SAL  $10^{-6}$  or better
- dosimetric monitoring
- microbial indicators

# Radiation

## $\gamma$ -radiating isotopes: (Ph.Hg.VII)

### **Cobalt-60 ( $^{60}\text{Co}$ ) $\rightarrow$ ( $^{58}\text{Co}$ in Ph.Eur 6, half time=70 days)**

Half-time: 5.3 years

it is produced by  $^{59}\text{Co}$  isotope with neutron-shooting

Eliminate  $\beta$ -radiation ( $> 0.3$  MeV) and  $\gamma$ -radiation (2 photons)  
(1,17 and 1,33 MeV)  $\rightarrow$  stable  $^{60}\text{Ni}$ -isotope

### **Caesium-137 ( $^{137}\text{Cs}$ )**

Half-time: 30 years

It is produced by uranium fission

1  $\gamma$ -photon is eliminated (0.66 MeV)



# Radiation

Lethal dose for different species

Species	Lethal dose (kJ · kg <sup>-1</sup> )
human	0.005 – 0.01
insects	0.01 – 0.25
bacteria	0.3 – 5.0
fungi	2.0 – 10.0
spores of bacteria	15.0 – 20.0
viruses	20.0 – 50.0

# Filtration

**Filtration:** adsorption and sieve effect

Sintered-glass filter

(G5): 1-1.5  $\mu\text{m}$   
sieve effect



Seitz-filters

EKS-1: 1.0-1.2  $\mu\text{m}$   
asbestos + cellulose  
adsorption



Membrane filters

0.20-0.45  $\mu\text{m}$   
cellulose polymers  
sieve effect





# Filtration

## Filtration (Ph.Eur. 6):

- product that cannot be thermally sterilized
  - „microbial challenge test” (*passage through the filter?*)

## Preparation!!!

- *The location of filtering and filling must be near to each other.*
- 0.22 µm pore size or less
- The solutions do not absorb on the filters.
- Avoid the contaminants from the filter
- Stage of filter = „bubble point” (tested before and after use)

## Attention is given to:

- **The bioburden prior to filtration**
- **Filter capacity**
- **Batch size**
- **Duration of filtration**

# Filtration

## Filtration (Ph.Eur. 6):

- asbestos fibers must be **eliminated**
- The membrane filters are made from **cellulose-nitrate** and -**acetate** or **polycarbonate**
- pore size of membrane filters= **0.20  $\mu\text{m}$** ,  
in case of viscous solution= **0.45  $\mu\text{m}$** .
- If an eye drop is filtered, than the preparation must be **preserved**.

# Filtration

## Bacteria-retentive membranes

- Fast and great volume
  - Surface area
  - Pressure
  - Suction
- By infusions
  - **Pre-filtration**
    - Glass-fiber filter
  - **Depyrogenation**
    - Activated carbon
  - **Bacteria-retentive membranes:**
    - Cellulose polymers (CA,CN)
    - 0.45 or 0.22  $\mu\text{m}$

# Gas sterilization

- Is only to be used where there is **no suitable alternative**
- Essential the penetration of gas and moisture into the materia
- End of the process: **residuals should not remain**
- Concentration, humidity, temperature, duration (register!)
- Microbial indicators
- Sterility test must be made from each batches

# Gas sterilization

## Chemical agents:

if the materia is heat-sensitive

formaldehyde,  $\beta$ -propiolactone, ethylen-oxyde



Equipments and textiles

### Drawback:

- Low penetration
- Odour, toxic
- Heavy removing (steam-rinsing)
- Polymerisation-coagulation



Non-irritable  
Less-volatile  
Effective in low concentration

### Drawback:

- Penetration into porous materia is bad
- Above 50°C polymerisation.



Effective  
Good penetration capability,  
Effective in room-temperature

### Drawback:

- It is used in the suitable gas mixture
- The moisture of the materia can reduce its effectiveness.

# Plasm sterilization

## With hydrogen-peroxide:

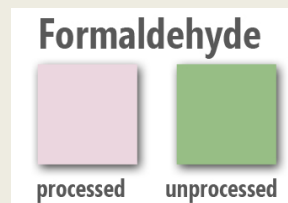
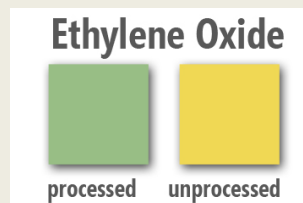
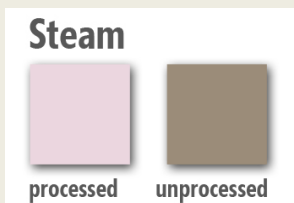
Vacuumed workplace – spray the hydrogen-peroxide – irradiate with radio-wave (electromagnetic) – cold-plasm state (ions, free radicals)

- medical equipment, metal tools, hand tattoo equipment, radiationtherapy devices, ultrasound heads
- Warning: cellulose, paper, liquids, powders, (vacuum)
- Temperature of sterilization:  $46 \pm 4$  °C
- Ventillation: trough HEPA-filter (pressure equalization)
- Cyclical period: 28 min – 1 hour

# Indicators: physico-chemical / biological

## Indicators

- **Indicator tape**
- **Indicator strips**  
(*non-uniform color = air in the workspace*  
not colored spots – overheated steam)
- **Brown's indicator tube**
- **Bowie Dick indicator pack**  
(*appearance of: ventilation, air infiltration, influx of non-condensable gases into the steam*)



# Heat indicator





# Biological indicators of sterilization (Ph. Eur. 6)

- „**Biological indicators are standardized preparations of selected micro-organisms used to assess the effectiveness of a sterilization procedure.**”
- „They usually consist of a population of bacterial spores placed on an inert carrier, for example a strip of filter paper, a glass slide or a plastic tube.”
- „may be inoculated directly into a liquid product to be sterilized.”

# Biological indicators


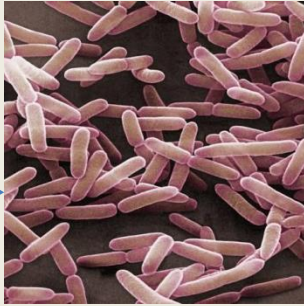
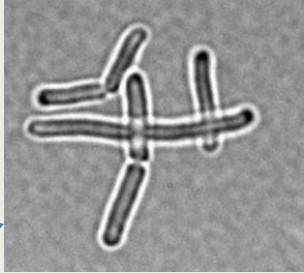
- Name of the species of the bacterium
- The number of the strain in the original collection
- Number of viable spores per carrier
- D-value (value of a parameter of sterilization required to reduce the number of viable organisms to 10 per cent of the original number)
  
- „The resistance of the test strain to the particular sterilization method is great compared to the resistance of all pathogenic micro-organisms and to that of micro-organisms potentially contaminating the product.”
  - „The test strain is non-pathogenic”
  - „The test strain is easy to culture”

# Biological indicators

- „It is recommended that the indicator organisms are placed at the locations presumed, or wherever possible, found by previous physical measurement to be least accessible to the sterilizing agent.”
- „After exposure to the sterilizing agent, aseptic technique is used to transfer carriers of spores to the culture media, so that no contamination is present at the time of examination.”
- „After incubation, growth of the reference micro-organisms subjected to a sterilization procedure demonstrates that the procedure has been unsatisfactory.”
- Species:
  - *Bacillus stearothermophilus*
  - *Bacillus subtilis*
  - *Bacillus pumilus*

# Biological indicators

Methods	Micro-organisms
Steam	B. Stearothermophilus
Dry heat	B. Subtilis var niger
Gas	B. Subtilis var niger
Radiation	B. pumilus



The table lists biological indicators for different sterilization methods. Blue arrows point from the 'Micro-organisms' column to corresponding micrographs on the right. The micrographs show: 1) B. Stearothermophilus as several long, thin, rod-shaped bacteria. 2) B. Subtilis var niger as a dense population of shorter, thicker rod-shaped bacteria. 3) B. pumilus as a dense population of very short, thick rod-shaped bacteria.

# Aseptic compounding and dispensing

- „The objective of aseptic processing is to **maintain the sterility of product that assembled from components.**” (Ph.Eur. 6)
- **Aseptic processing:** the whole of those work methods and processes – including the person’s behavior and health status – which can keep away the microorganisms.
- „This is achieved by using conditions and facilities designed to prevent microbial contamination.”

# Aseptic preparation

## Careful attention:

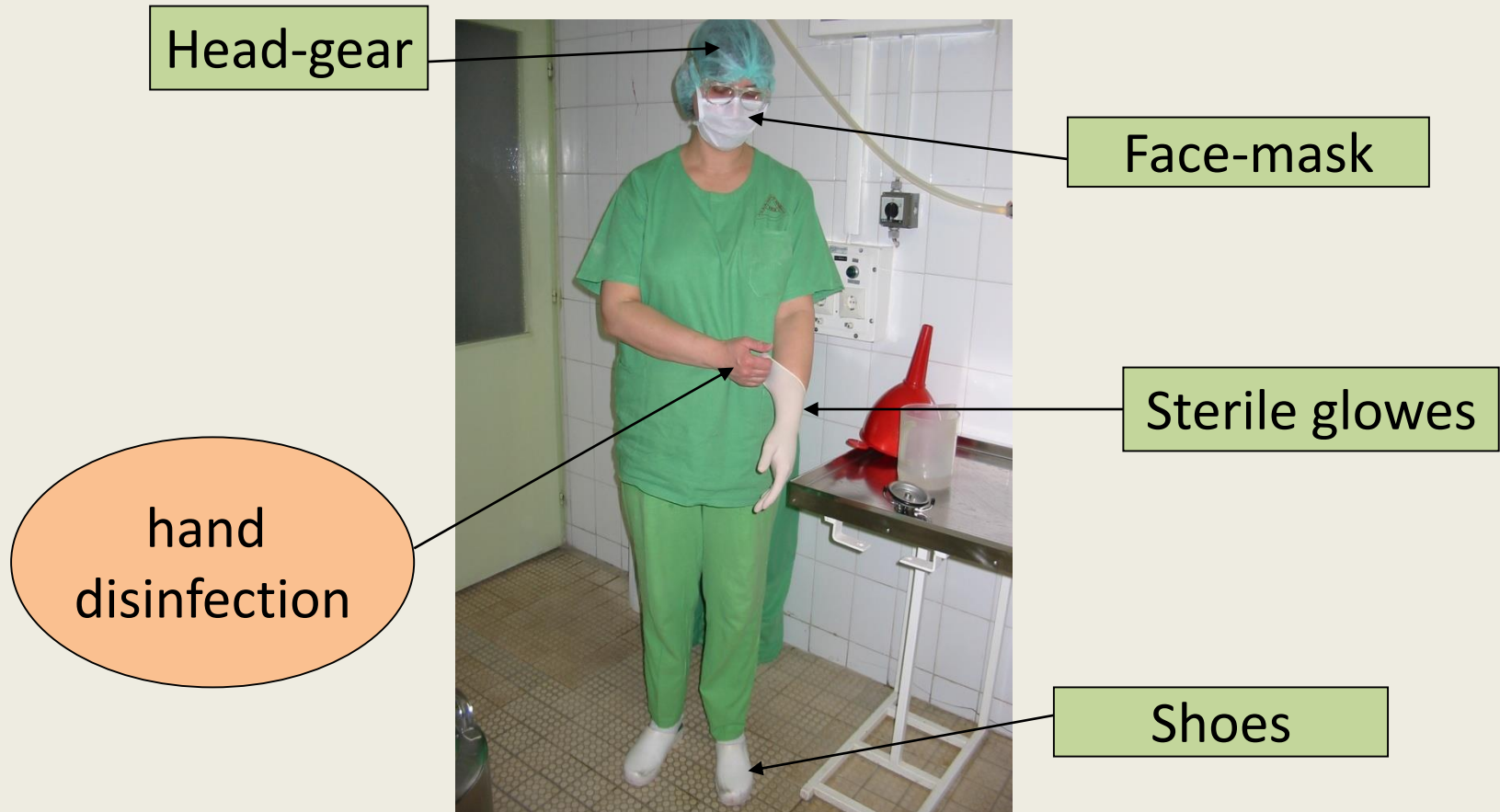
- environment
- personnel
- critical surface
- container/closure sterilization and transfer procedure
- maximal holding period of the product before filling into the final container

# Aseptic preparation



Appropriate concentration and time (with new solution)

# Aseptic preparation





# Aseptic preparation

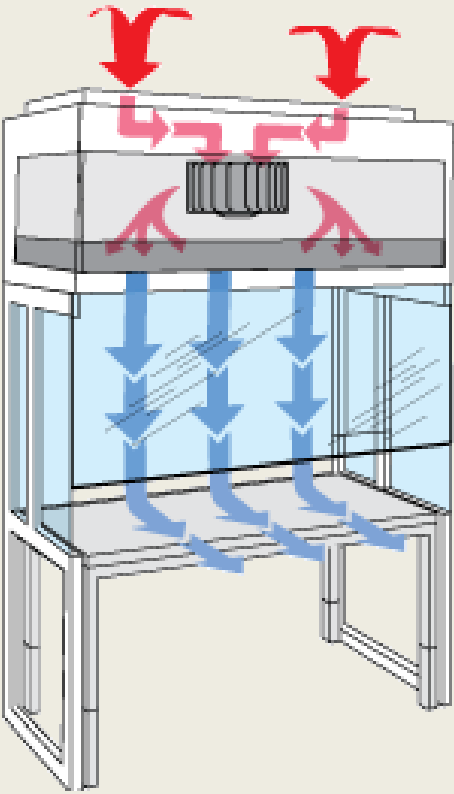
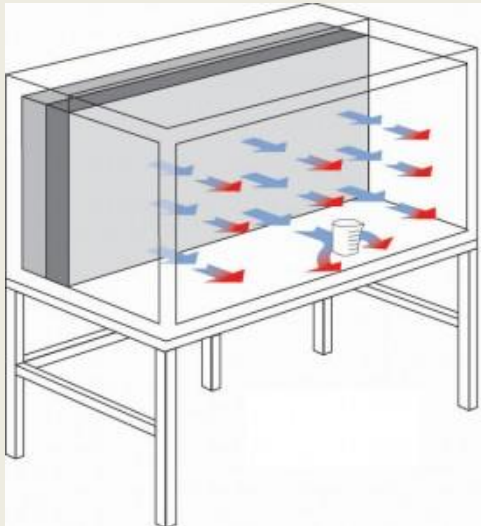


# Manipulator

UV-radiation (Germicide)



# Laminar box



# Aseptic preparation

Infusion bottle



- cleaning
- rinsing with distilled water
- sterilizing

rubber stoppers



# Sterilization

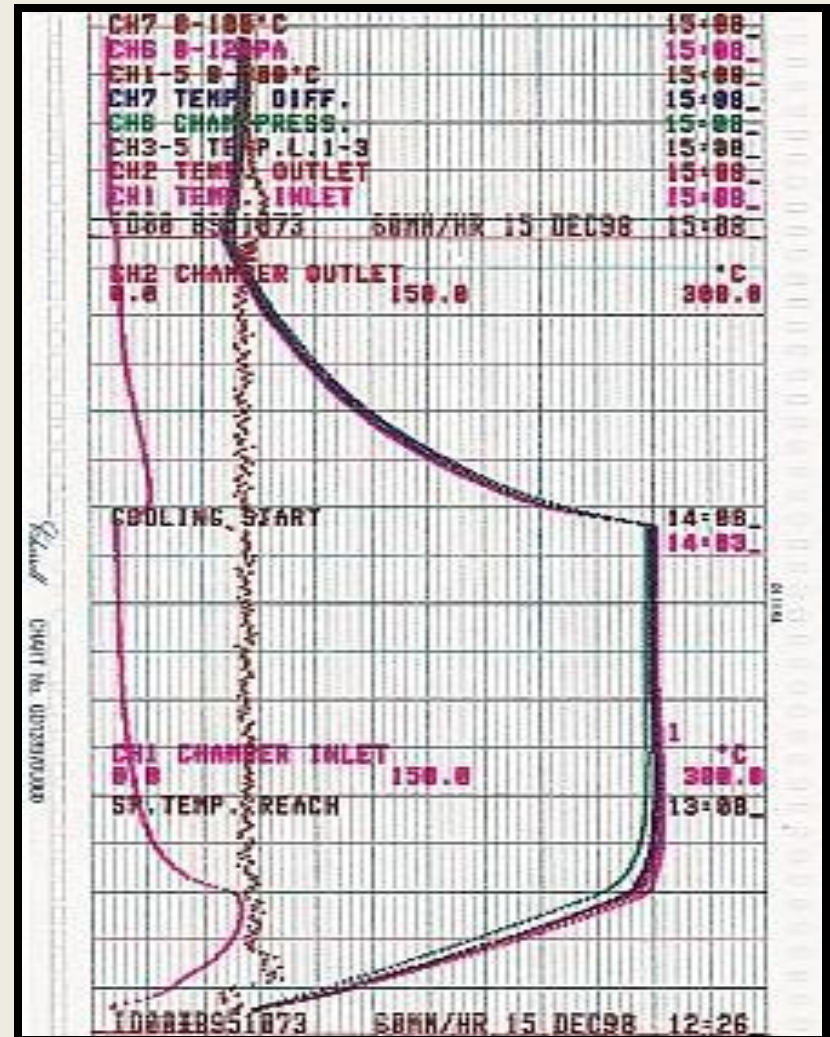
## Controlling:

- pressure
- temperature
- indicator
- chemical
- microbiological

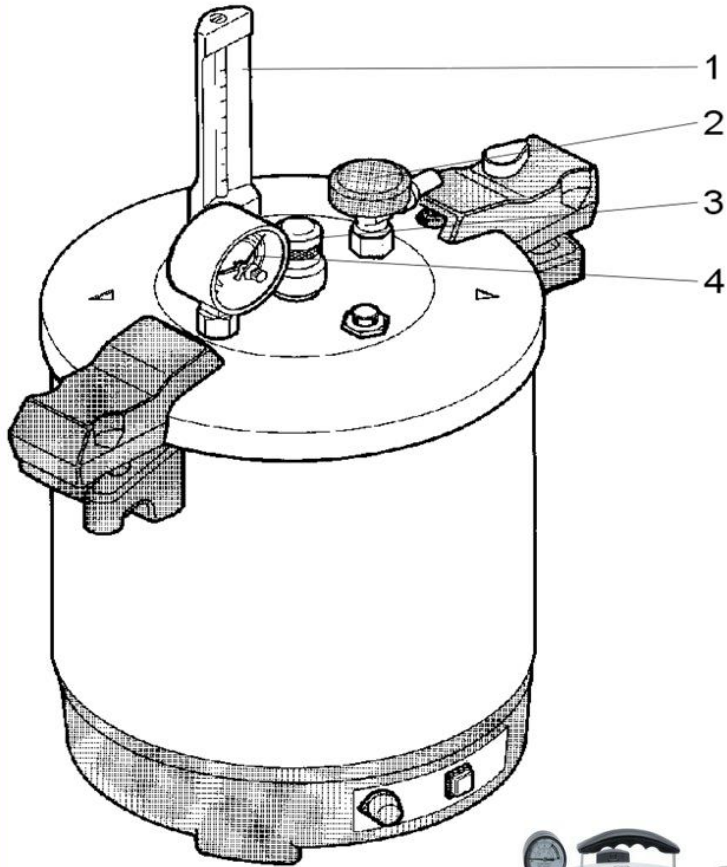


# Sterilization

## *Process controlling*

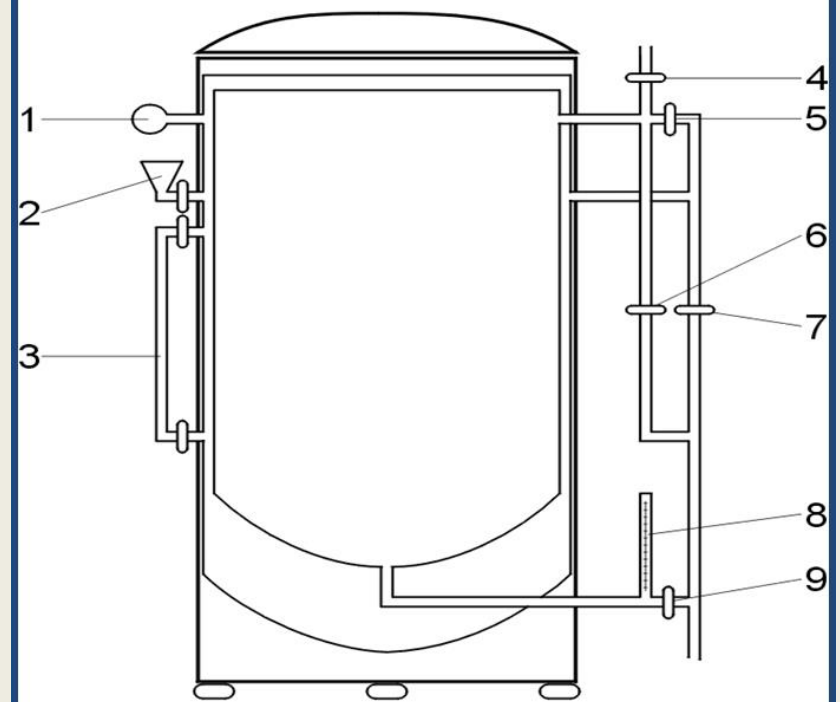


# Certoclave and autoclave



## Certokláv

1. Thermometer
2. Steam drain tap
3. Pressure control valve
4. Pressure gauge



## Autokláv

1. Ventillation valve
2. Water inlet
3. Water level indicator
4. Air suction tap
5. Steam inlet tap
6. Vacuum valve
7. Vacuum valve
8. Thermometer
9. Condensation tap



# Depyrogenation

- **Pyrogens: materials, that can cause fever** (endo-, exogen), coagulation disorders and metabolic changes.
- Endotoxins of Gram-negative bacterias (lipopolysaccharide (LPS)) G-80%, G+20%,
- It binds to the external membrane
- When the bacterium dies, LPSs can dissolve into the media.
- Pyrogens must not be included in parenteral (>50ml) preparations.



## Biological effect of endotoxins:

- fever
- coagulation
- metabolic changes



# Depyrogenation

- Pyrogen's chemical resistance is very high
- 200-250°C temperature is needed to destroy them
- Sensitive to oxydation
- It can be adsorbed by adsorbents:
  - **active carbon**
  - asbestos-filter

# Examinations to pyrogens (Ph.Eur.6)

In vitro: **LAL-test** (**L**imulus **A**moebocyta **L**isatum)

„The test for bacterial endotoxins is used to detect or quantify endotoxins of G- bacterial origin using amoebocyta lysate from horseshoe crab (*Limulus polyphemus*).”

- gel-clot technique (based on gel formation)
- Turbidimetric technique (based on the development of turbidity after cleavage of an endogenous substrate)
- Chromogenic technique (based on the development of colour after cleavage of synthetic peptic-chromogen complex.)

They may be kinetic, semi-quantitative or end-point methods.

In vivo: **rabbit pyrogen test**

Inserted into veins of rabbit's ears.

Detecting the body temperature.

# LAL-test

## Advantage:

- In vitro process
- fast, duration: 1.5 h
- sensitivity is good
- cheap

## Disadvantage:

- just G -, (no virus)
- interaction with  
( $\text{HCO}_3$ , phosphates, citrates, glucose)
- It is water based test. (oil based hormones!!)
- inorganic materials (rubbers, closures) are not measurable.

 BIO



Limulus ameocyte lysate (LAL) is an aqueous extract of blood cells (amoebocytes) from the blue-blooded horseshoe crab, *Limulus polyphemus*. LAL is being used for endotoxin detection.

# Sterile preparations

- Injections
- Infusions
- Single dose eye drops, eye ointments
- Powders for tablets for parenteral administration
- Implants
- Enteral and parenteral nutrition
- Haemodialysis solutions
- Aerosols for inhalation and wound surfaces
- Other dosage forms (wound ointment / gel, wound sprinkling powder)

# Ph.EUR. (Ph.Hg.VIII.)

Micro-biological class	Preparations	Requirements
1.	Sterilization is required by the appropriate dosage form and sterile according to the label of other preparations	<b>Sterility</b>

# Ph.EUR. (Ph.Hg.VIII.)

Micro-biological class	Preparations	Requirements
2.	Topical and respiratory pharmaceutical preparations, except where sterility is required Transdermal patches (Tests both adhesive and substrate)	all viable $10^2$ aerobic bacteria and fungi / g or ml -allowed: $10^1$ enterobacteriaceae and other gram-negative bacteria / g or ml - except: not for transdermal patches! excluded: Pseudomonas aeruginosa (1.0 g or 1.0 ml) Staphylococcus aureus (1.0 g or 1.0 ml)

# Ph.EUR. (Ph.Hg.VIII.)

Micro-biological class	Preparations	Requirements
3.	<p>A. / Pharmaceuticals for oral and rectal administration</p> <p>B. / Oral preparations of natural (animal, plant, mineral) origin, for which antimicrobial pre-treatment is not possible and whose starting material is <math>10^3</math> viable micro-organisms / g or ml from the competent authority. Herbs included in microbiological class 4 are excluded.</p>	<p>Total viable aerobic count <math>10^3</math> bacteria/g or ml and <math>10^2</math> fungi/g or ml</p> <p>Excluded:</p> <p>Escherichia coli (1.0 g or 1.0 ml)</p> <p>Total viable aerobic micro-organism count <math>10^4</math> aerobic bacteria/g and ml <math>10^2</math> fungi/g or ml</p> <p>Not more than <math>10^2</math> enterobacteriaceae and other gram-negative bacteria / g or ml</p> <p>excluded:</p> <p>Salmonella (10.0 g or 10.0 ml)</p> <p>Escherichia coli (1.0 g or 1.0 ml)</p> <p>Staphylococcus aureus (1.0 g)</p>

# Ph.EUR. (Ph.Hg.VIII.)

Micro-biological class	Preparations	Requirements
4.	<p>This class includes herbal preparations containing exclusively herbal, whole, comminuted or powdered drug (s).</p> <p>A ./ Herbal preparations that need to be boiled before use.</p> <p>B. / Herbal preparations that do not require scalding before use.</p>	<p>Total viable aerobic micro-organism count: <math>10^7</math> aerobic bacteria/g or ml and <math>10^5</math> fungi/g or ml</p> <p>Not more than 10 <i>Escherichia coli</i>/g or ml</p> <p>Total viable aerobic count <math>10^5</math> aerobic bacteria/g or ml and <math>10^4</math> fungi/g or ml</p> <p>Not more than <math>10^3</math> <i>Enterobacteriaceae</i> and other gram-negative bacteria/g or ml</p> <p>Excluded:</p> <p><i>Escherichia coli</i> (1.0 g)</p> <p><i>Salmonella</i> (10.0 g)</p>



**Thank you  
for your attention!**