

# Filtration, sedimentation, centrifugation,

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*Institute of Pharmaceutical Technology and Biopharmacy*



**BASIC OPERATIONS**

**FILTRATION**

# Filtration

**Filtration** is an **operation** (separation process), in which a **heterogeneous mixture can be separated** to its different forms of components (solid, liquid, gas).

Filtration is applied in case of:

- crystallization
- fiber examination of injections
- cleaning of air

# Filtration

The **driving forces** of filtration:

- gravitation,
- pressure or suction,
- centrifugal force.

# Filtration

Gravity force

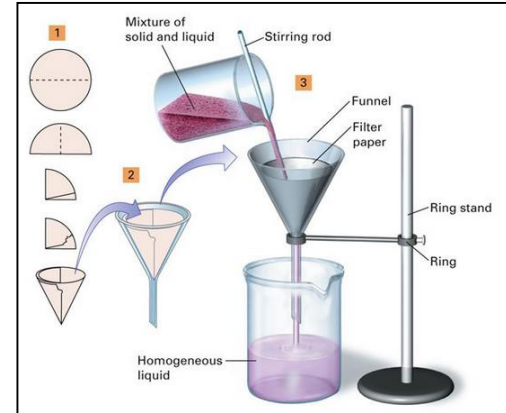
$F_g$

mixture

filter

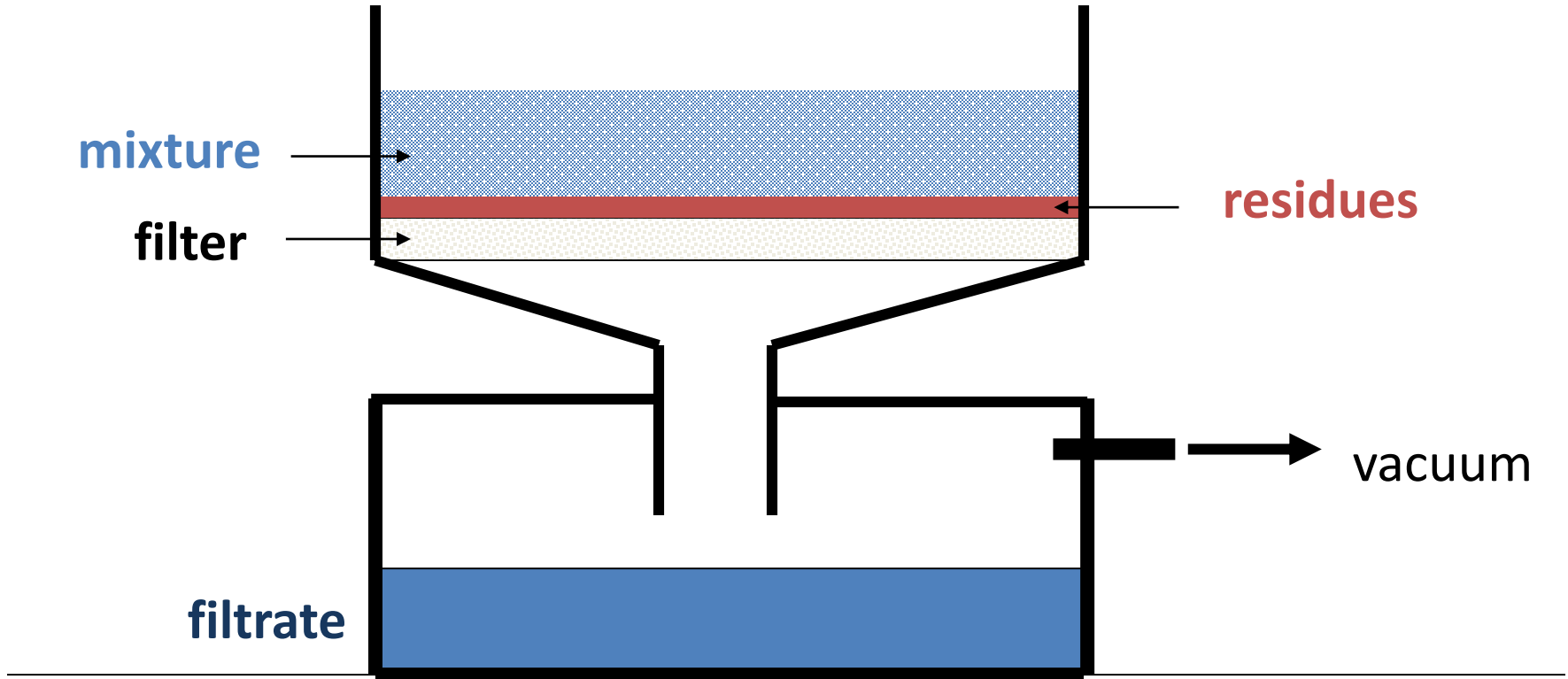
residues

filtrate



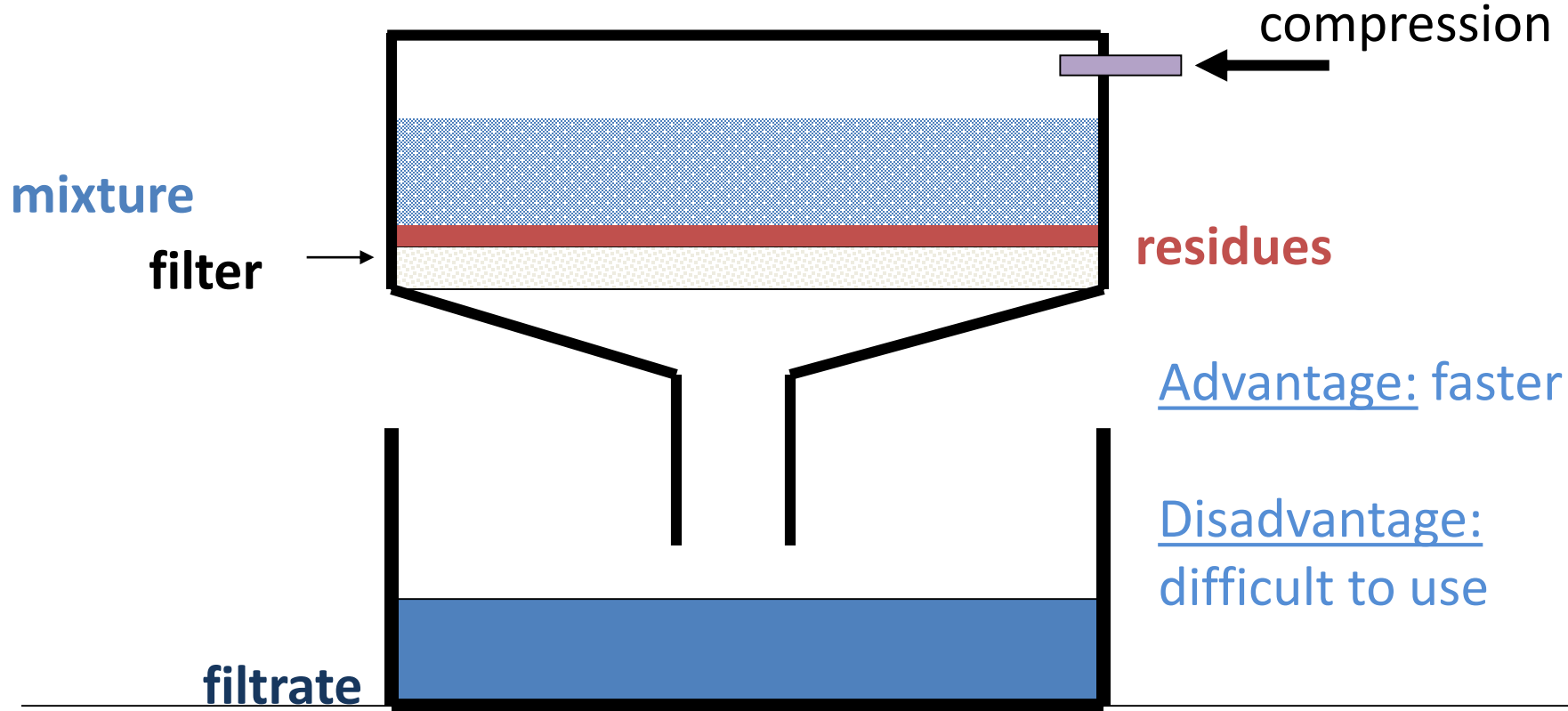
# Filtration

**Vacuum (suction)**



# Filtration

**Pressure**



compression

mixture

filter

residues

Advantage: faster

Disadvantage:  
difficult to use

filtrate

# Filtration rate

The filtration rate depends on:

- thickness of the bed
- permeability



# Filtration rate

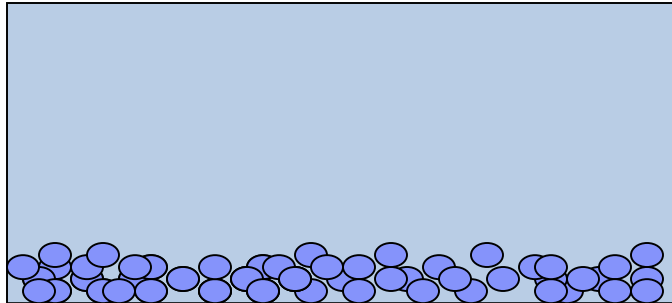
The residue may be

- **Incompressible**
  - the size of pores and canals is constant,
- **Compressible**
  - the size of pores and canals is decreasing during the filtration.

# Filtration rate

## Structure of the residue (1):

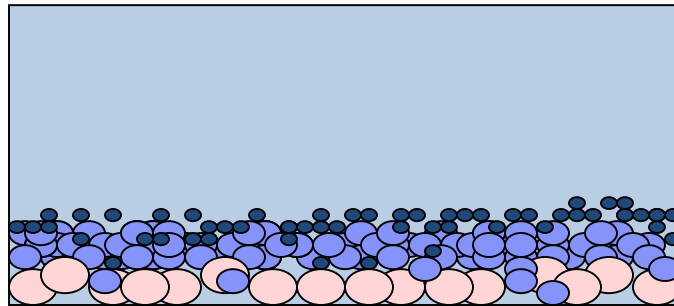
- homodisperse,
- the filtration rate is constant,
- no compaction.



# Filtration rate

## Structure of residue (2):

- heterodisperse
- the filtration rate is decreasing during the process,
- compaction



# Filtration rate

## Darcy equation

$$\frac{dV}{dt} = \frac{BA\Delta p}{\eta L}$$

$V$  = volume of the filtrated mixture

$t$  = time

$B$  = permeability

$A$  = area of the filter

$\Delta p$  = difference of the pressure

$\eta$  = viscosity

$L$  = thickness of the filter

# Filtration rate

## Kozeny-Carman equation

(fibrous filter media)

$$B = \frac{d_f^2 \cdot \varepsilon^3}{16(1 - \varepsilon)^2 k}$$

$B$  = permeability

$d_f$  = filter diameter

$\varepsilon$  = porosity

$k$  = Kozeny-Carman-constant

# Filtration rate

## Meyer-Smith equation

(granular filter medium )

$$B = \frac{d_{sp}^2 \cdot \varepsilon^{4,1}}{150(1 - \varepsilon)^2}$$

$B$  = permeability

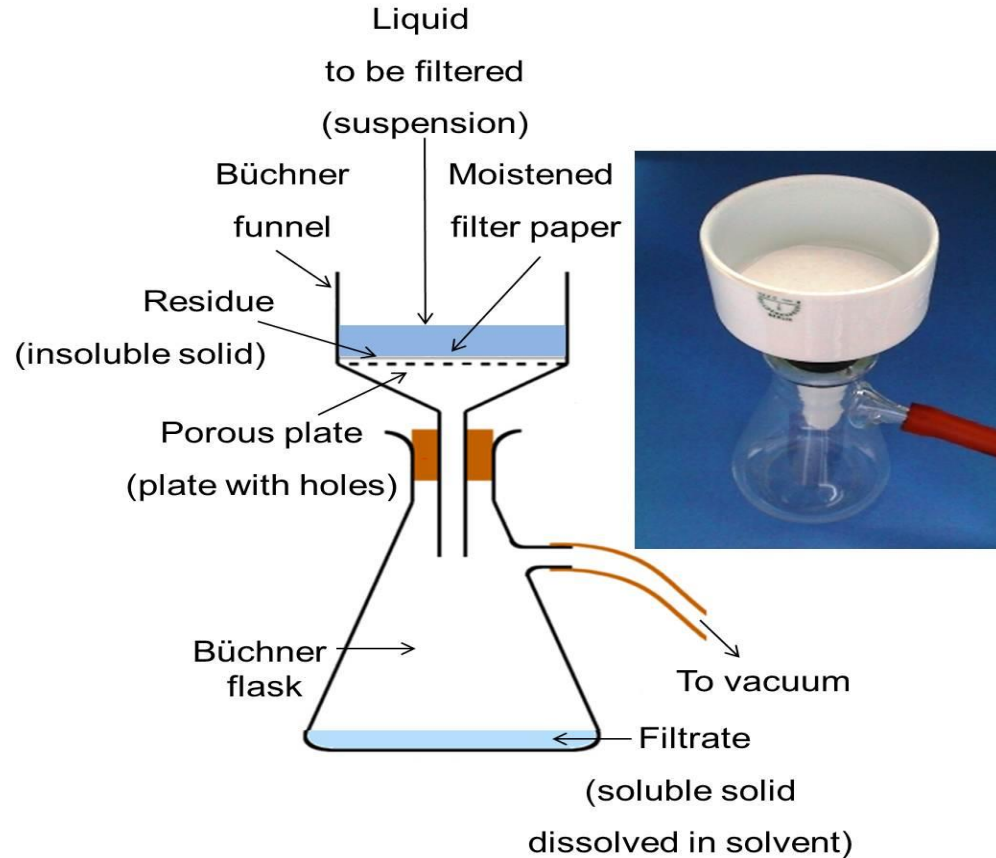
$d_{sp}$  = mean granule diameter

$\varepsilon$  = porosity

# Equipments

## Vacuum

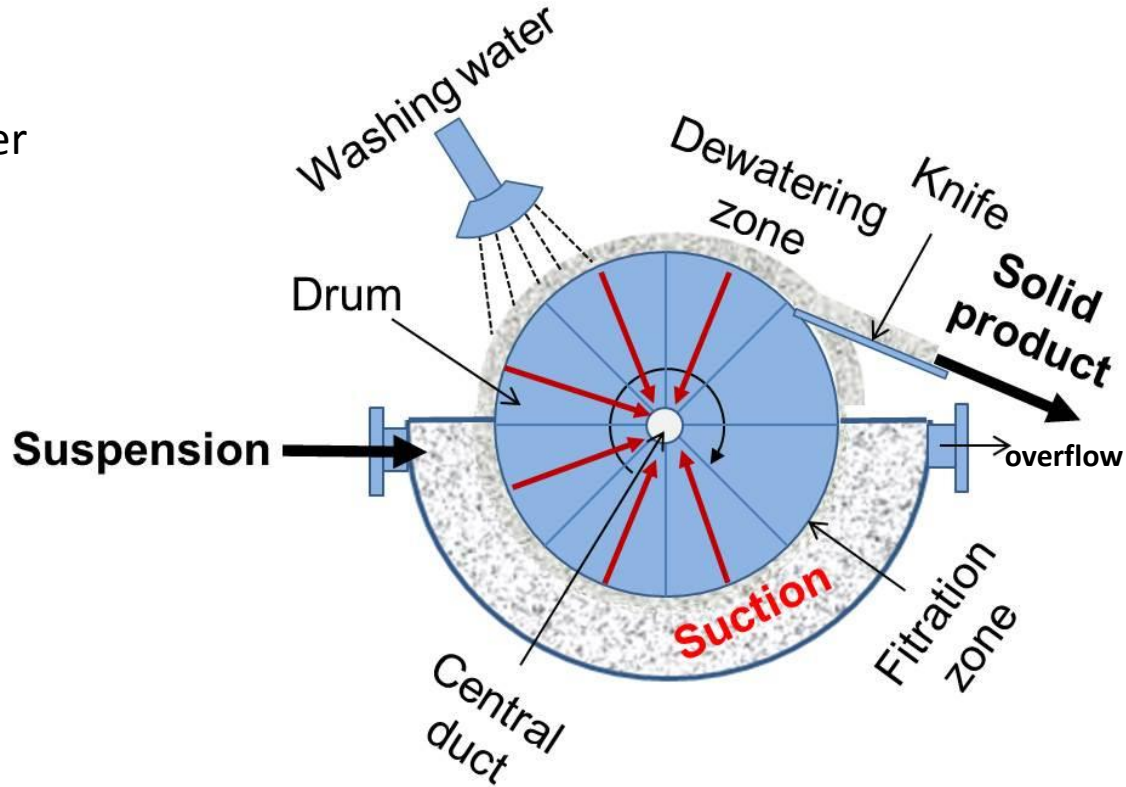
laboratory  
suction filter



# Equipments

## Vacuum

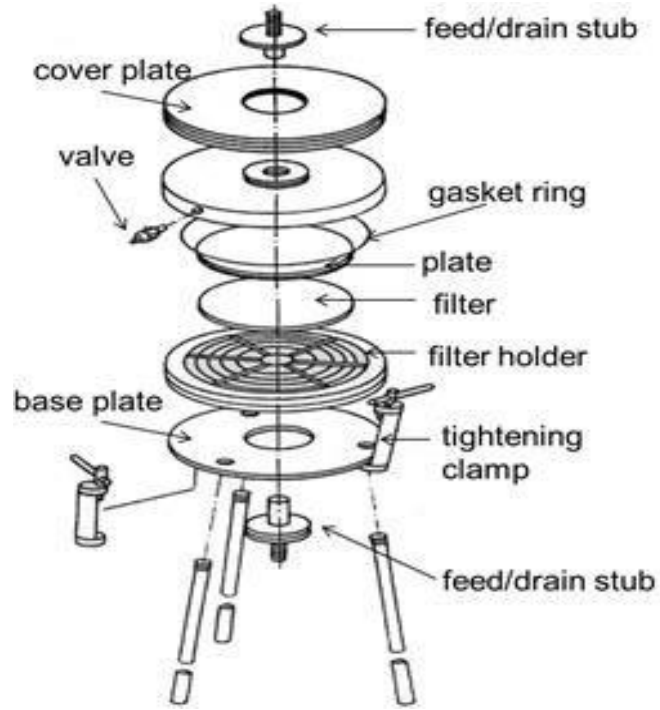
Vacuum  
rotary-drum filter





# Equipments

## Pressure

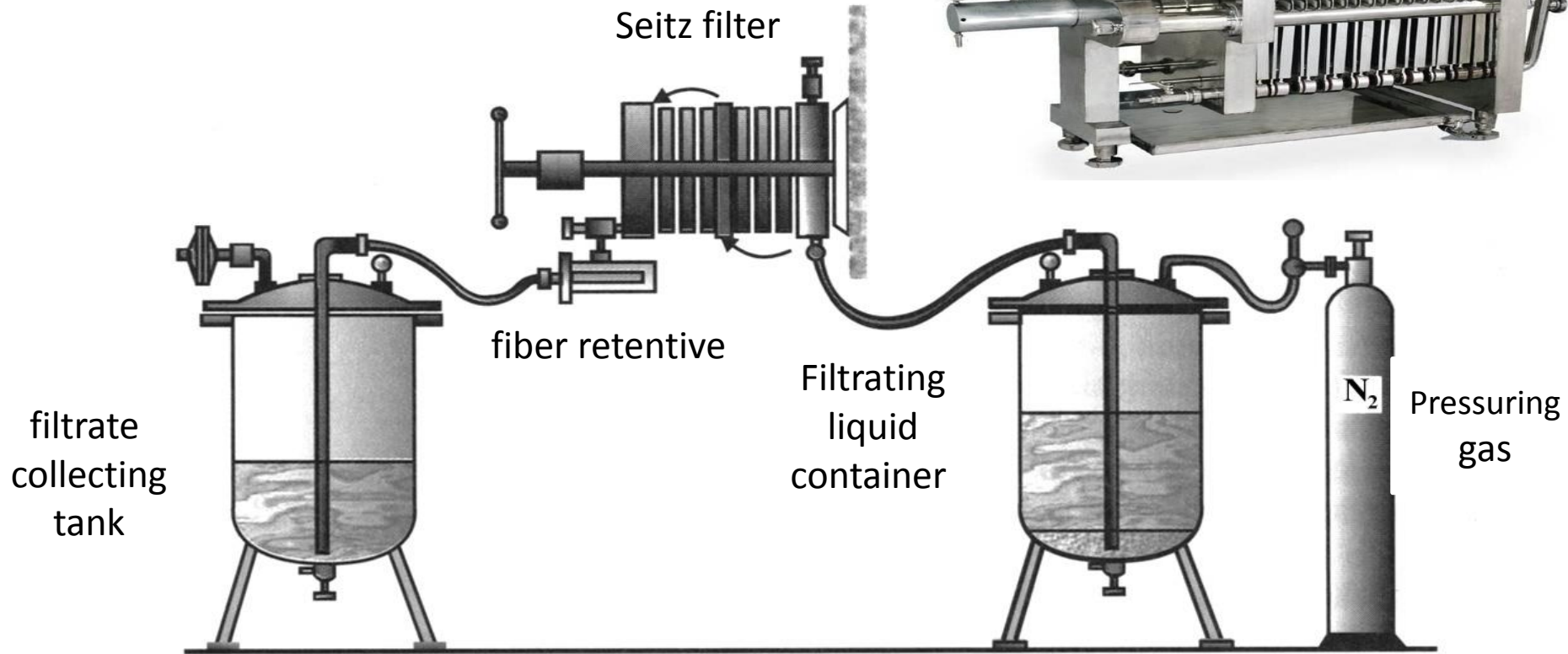


laboratory



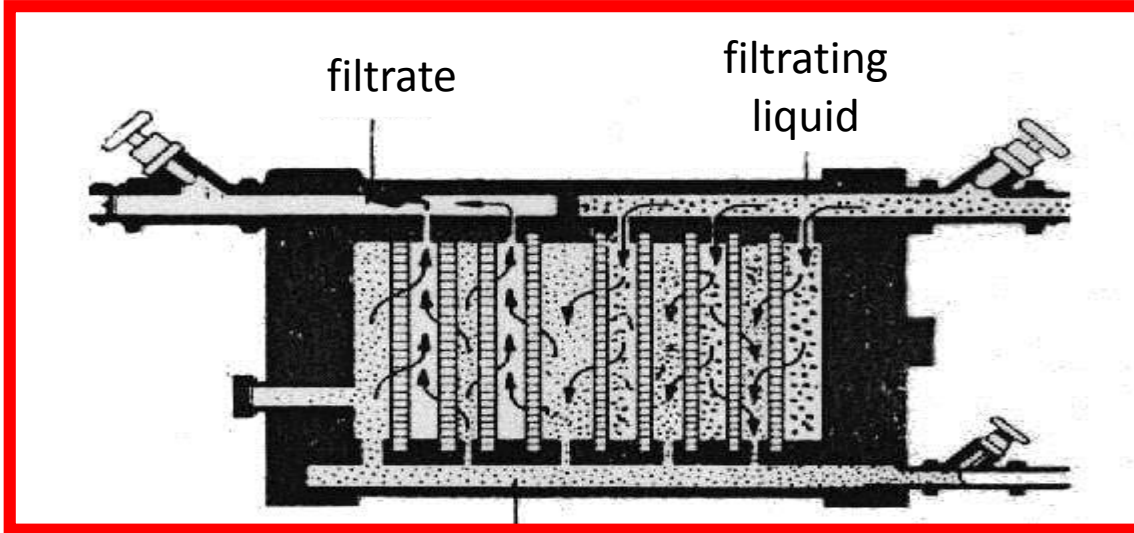
# Equipments

## Frame and plate filter

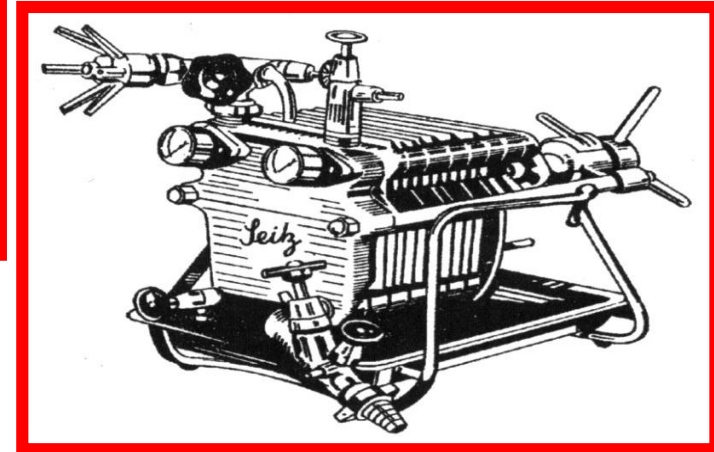


# Equipments

**Double filter press**

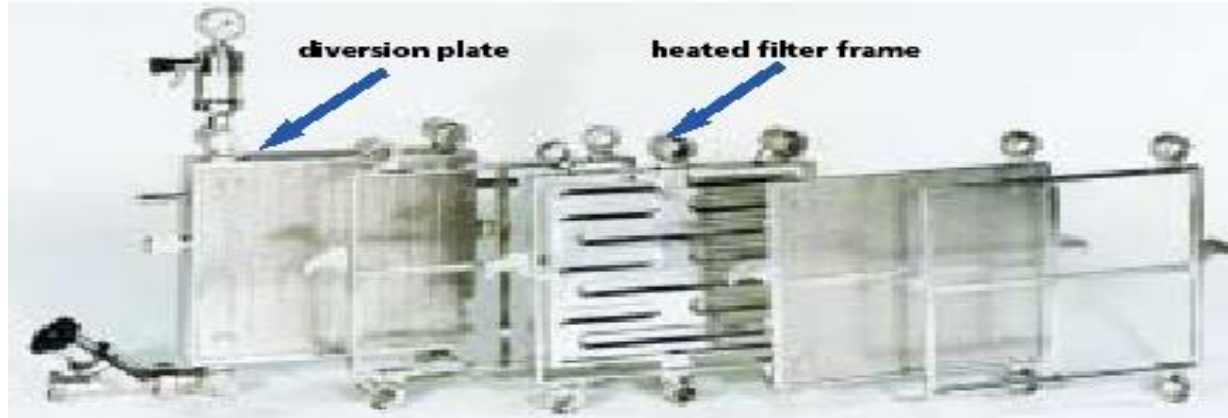


**frame filter**



# Equipments

## Frame filter



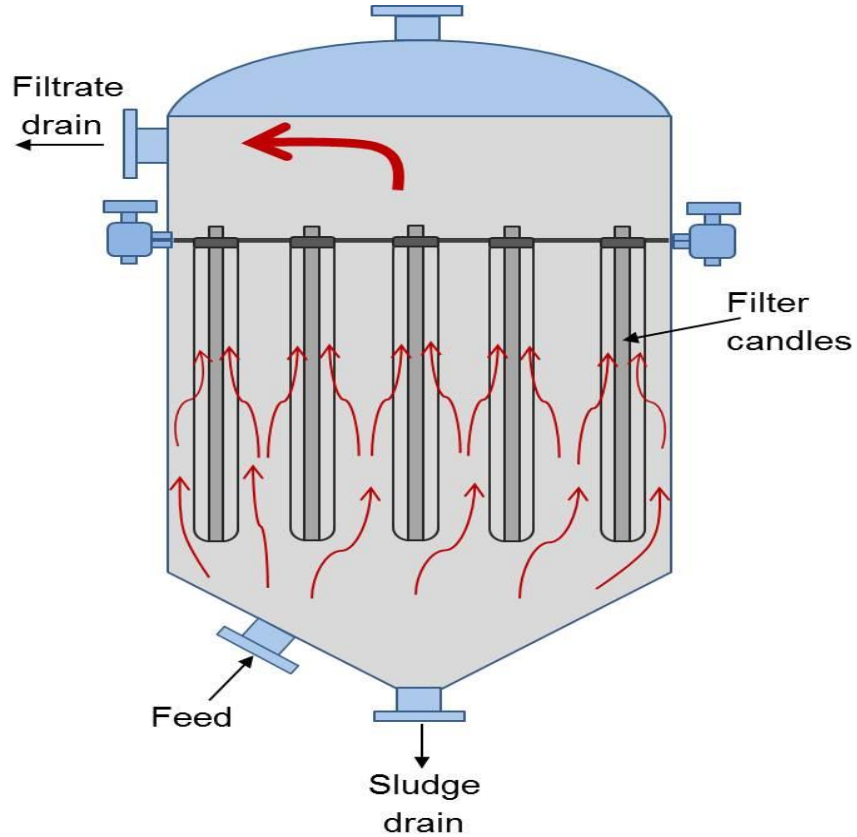
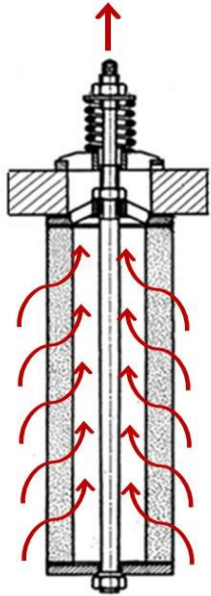
laboratory



industrial

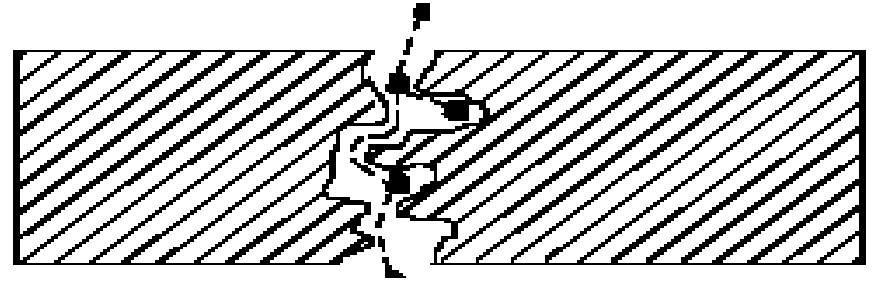
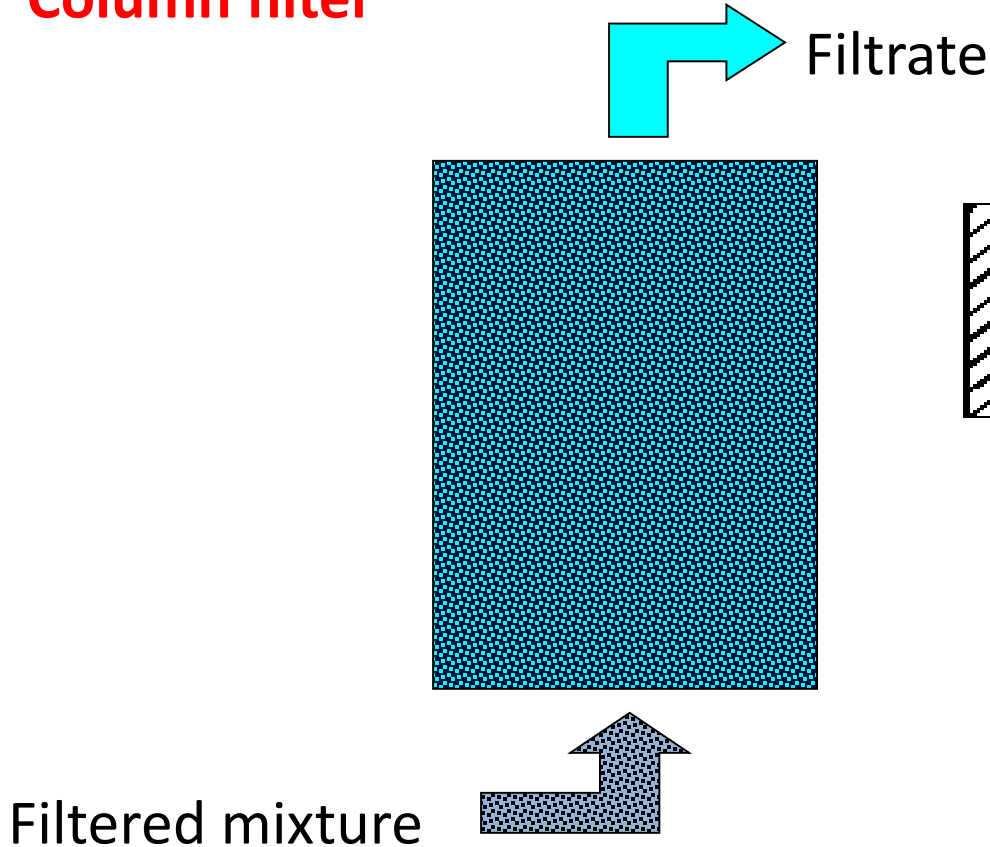
# Equipments

## Candle filter



# Equipments

## Column filter

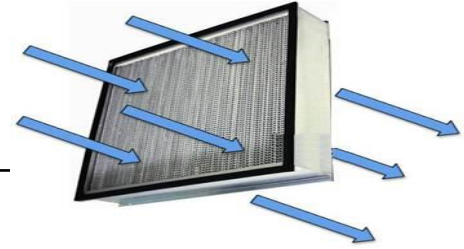


## capillar effect

Filled with granules or adsorptive substances (gravel, pellets)

# Seitz-filters

## Cellulose filters

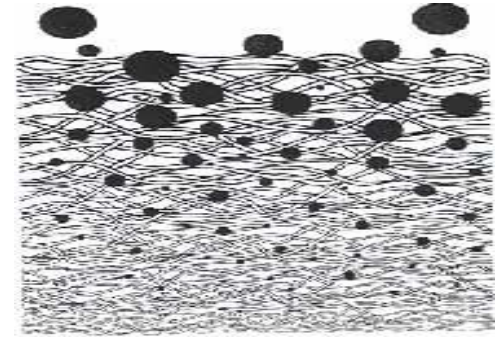
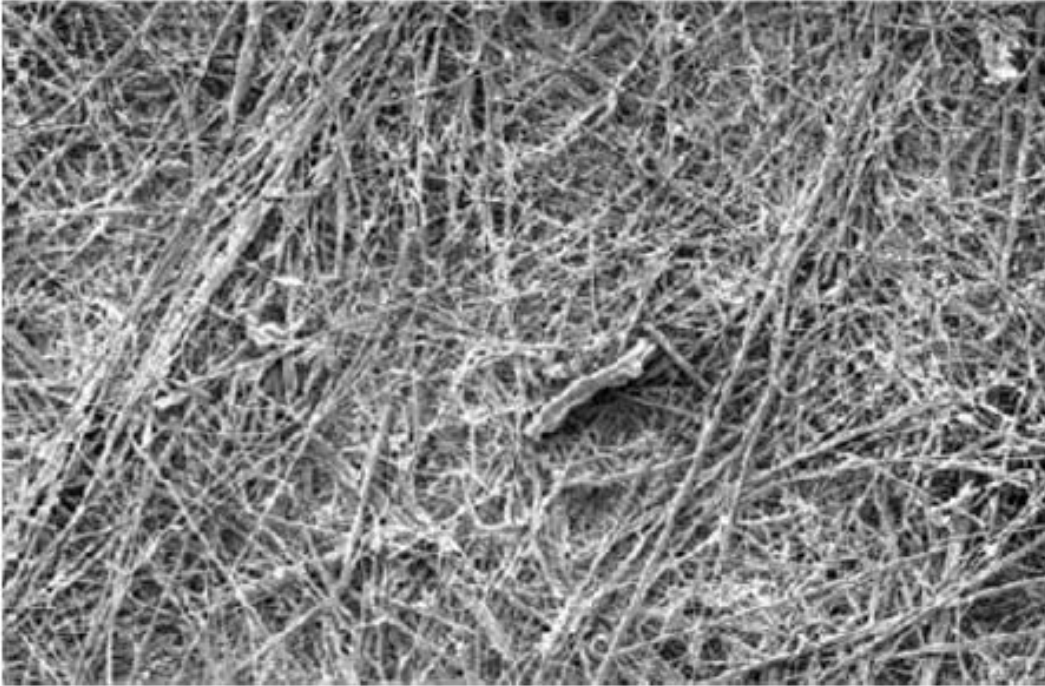


Types of Seitz-filter	Pore size ( $\mu\text{m}$ )	Filter capacity (ml/h)		Application
		19,62 kPa	49,05 kPa	
EK	1,4 – 1,8	200	500	<b>Germ-free drinks (galenical preparations, syrups, extracts)</b>
DKS	1,2 – 1,4	130	325	<b>Germ-, and pyrogen-free water, injections (aqueous), low molecular weight solution</b>
EKS-1	1,0 – 1,2	100	250	<b>Injections (aqueous), kolloidal solutions or higher molecular weight solutions (proteins), blood replacement</b>
EKS-11	0,8 – 1,0	75	190	<b>See: EKS-1, especially by blood and serums</b>



# Depth filters

## Depth (fibered) filters



**Filtration through the  
depth filter**



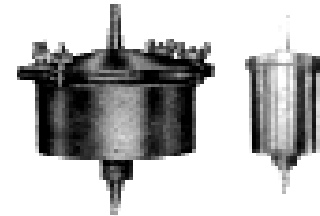
# Filters

## Rigid, porous filters

- ***Porous ceramic*** (Chamberlain filters, ~candles - kaoline  
Berkefeld filters - diatomaceous earth)  
clarifying and sterilizing filtration

- ***Glass filters*** ( Pyrex, Schott - sintered glass filter)

clarifying and fiber-exception and  
sterilizing filtration



# Properties of Chamberland filters

Types	Pore size (µm)	Application
Chamberland L 1	4,7 – 8,9	Clarify
Chamberland L 2	2,2 – 4,7	Clarify
Chamberland L 3	2,0 – 2,2	Clarify
Chamberland L 5	1,0 – 2,0	Sterilization
Chamberland L 7	1,0	Sterilization
Chamberland L 11	0,11	Sterilization

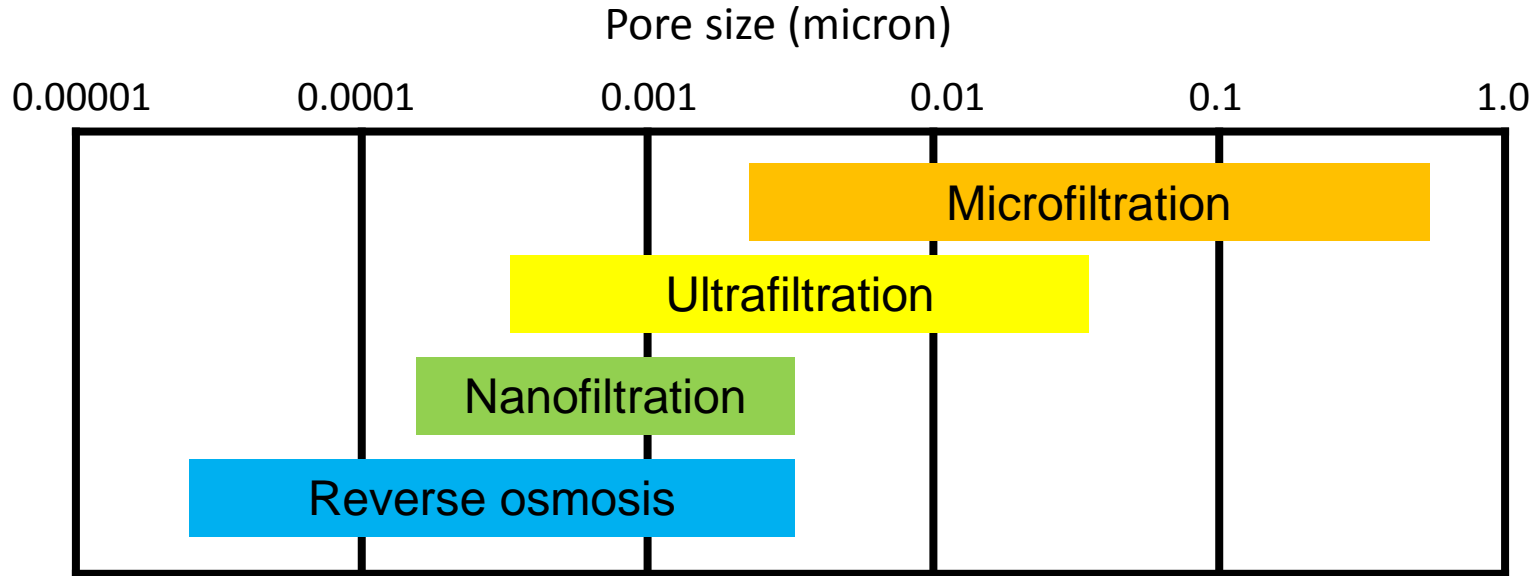


# Essential glass filter information

Type	Pore size (μm)	Application
G 00	200-500	refining solutions
G 0	150-200	refining solutions
G1	90-150	refining brews, infusions
G2	40-90	refining brews, infusions, syrups
G3	15-40	filtration of alcoholic and aqueous solutions
G4	3-13	filtration of eye drops
G5	1,0-1,5	sterilizing filtration of injection solutions

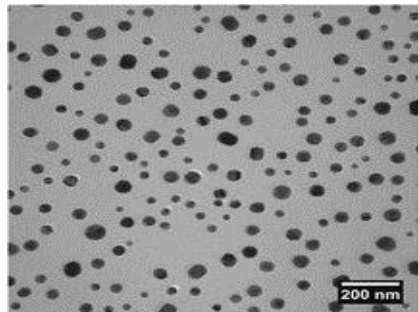


# Membrane filters

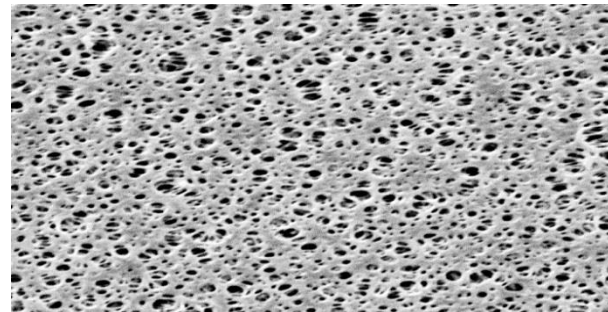
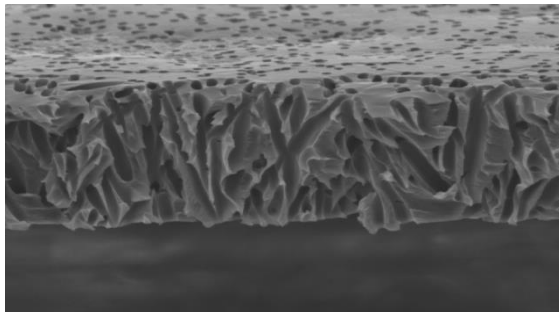


Typical pore sizes for membranes used in reverse osmosis, nanofiltration, ultrafiltration, and microfiltration.

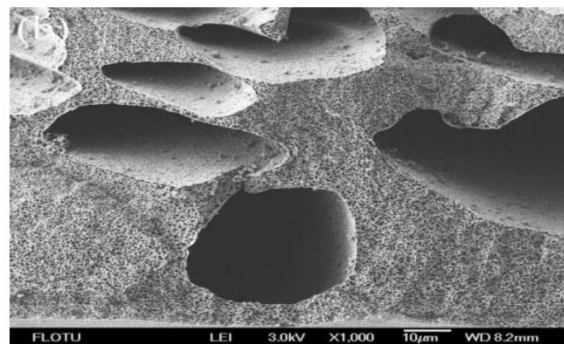
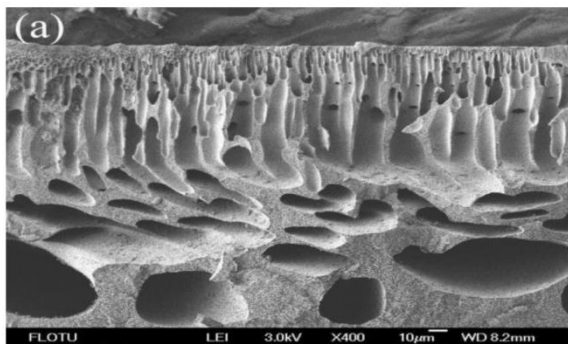
# Membrane filters



SEM of Polycarbonate membrane

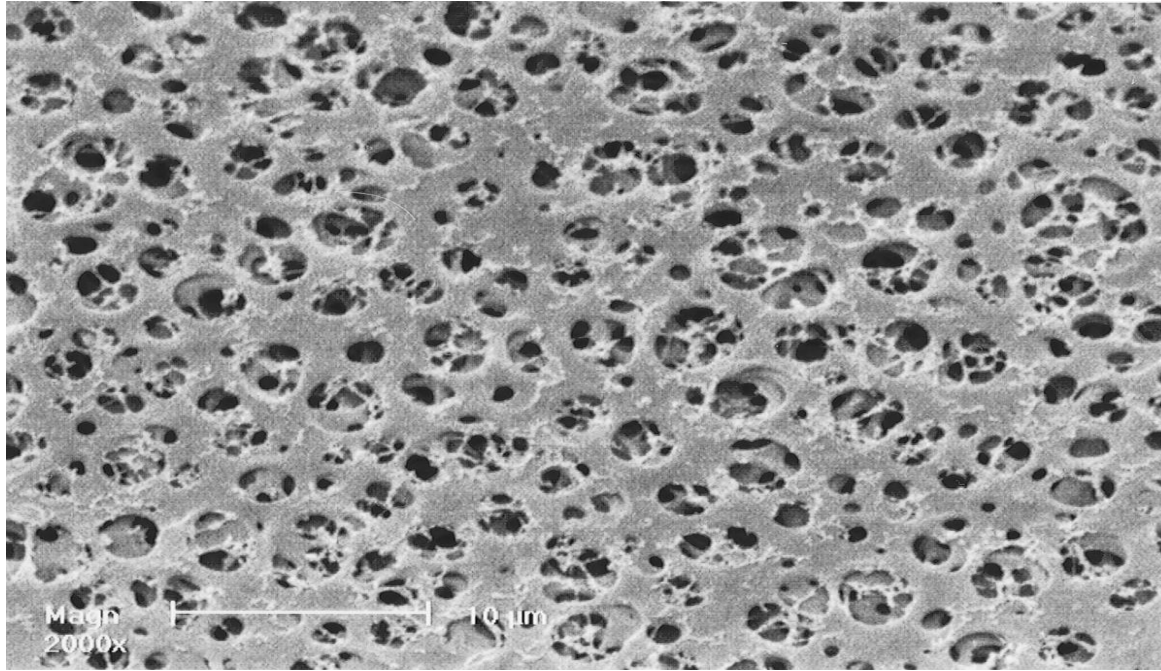


SEM of Cellulose  
(Polyethersulfone) membrane



Cross section of Polyphenylsulfone (PPSU) ultrafiltration membrane

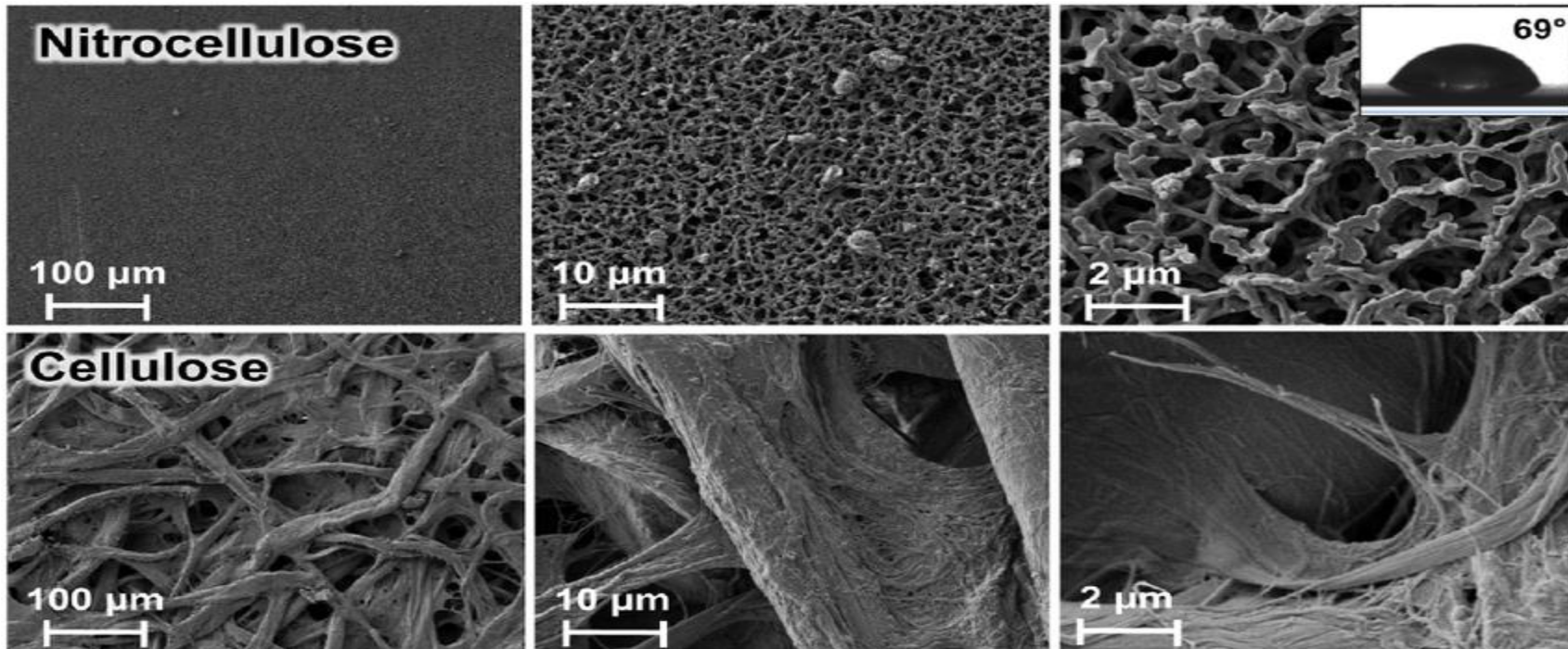
# Membrane filters



SEM of Cellulose acetate membrane

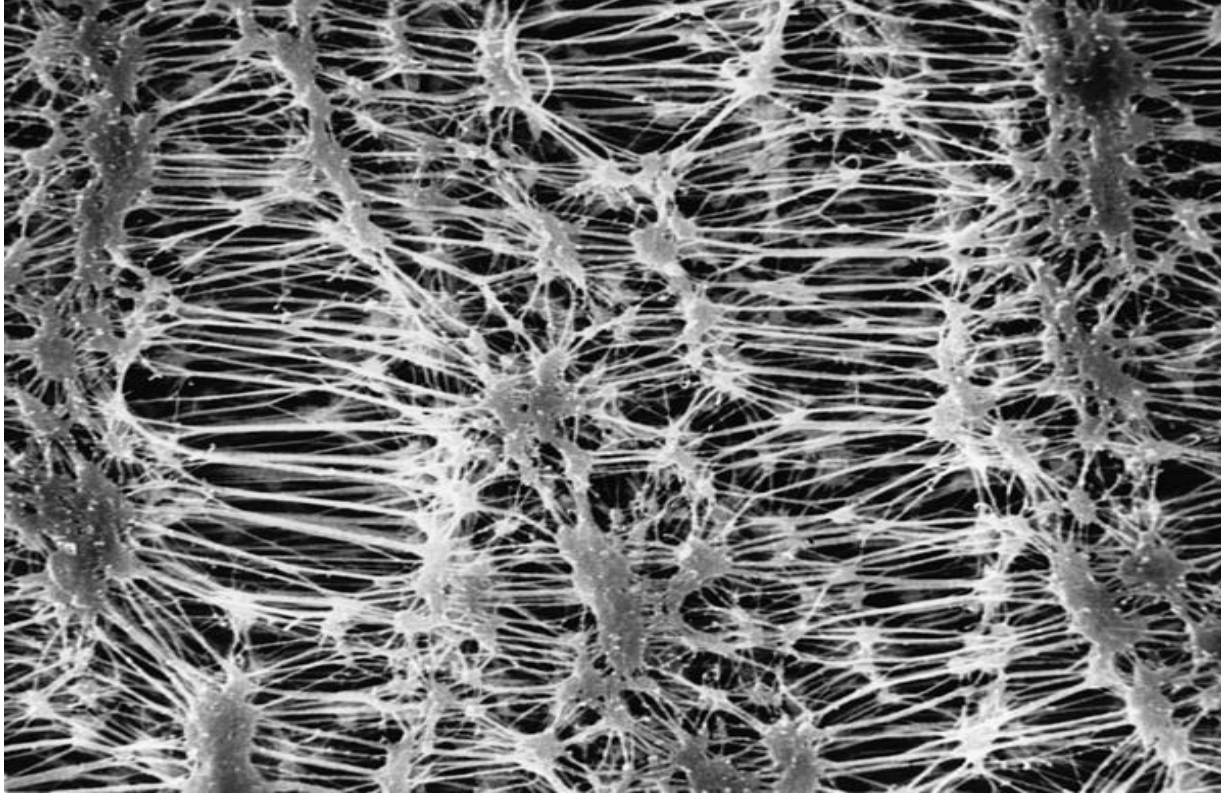


# Membrane filters



SEM of Nitrocellulose and Cellulose membrane

# Membrane filters



SEM of Fluoropore membrane



# Membrane filters

**Type**

**Manufacturer**

cellulose ester

Millipore

nylon

Duralon

cellulose ester and nylon

Mikroweb

cellulose acetate

Celolate

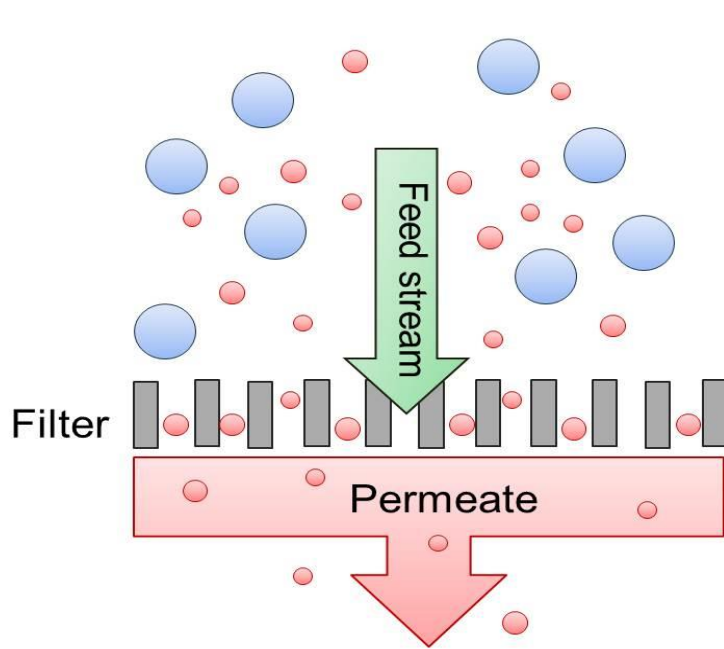
teflon

Mitex

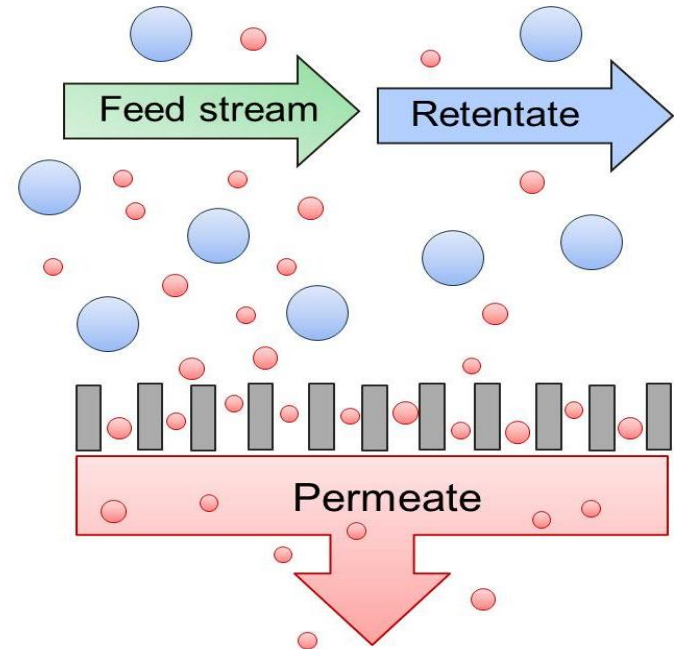
polyvinyl derivatives

Polyvic

# Main types of surface filtration



Dead end filtration



Cross flow filtration

# Membrane filters

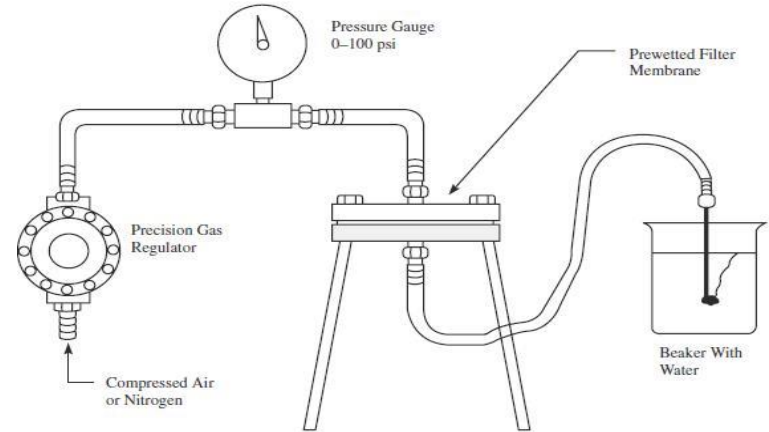
## Bubble Point Test

The bubble point measurement determines the **pore size of the filter membrane**, i.e., the larger the pore the lower the bubble point pressure.

At a certain pressure level, liquid will be forced first from the set of largest pore, in keeping with the inverse relationship of the

- *applied air pressure (P) and*
- *the diameter of the pore (d)*
- *$\gamma$  is the surface tension,*
- *$\vartheta$ =wetting angle:*

$$P = \frac{4\gamma \cos \Theta}{d}$$



The bubble point test can be associated with the result of the **bacterial retention** test.

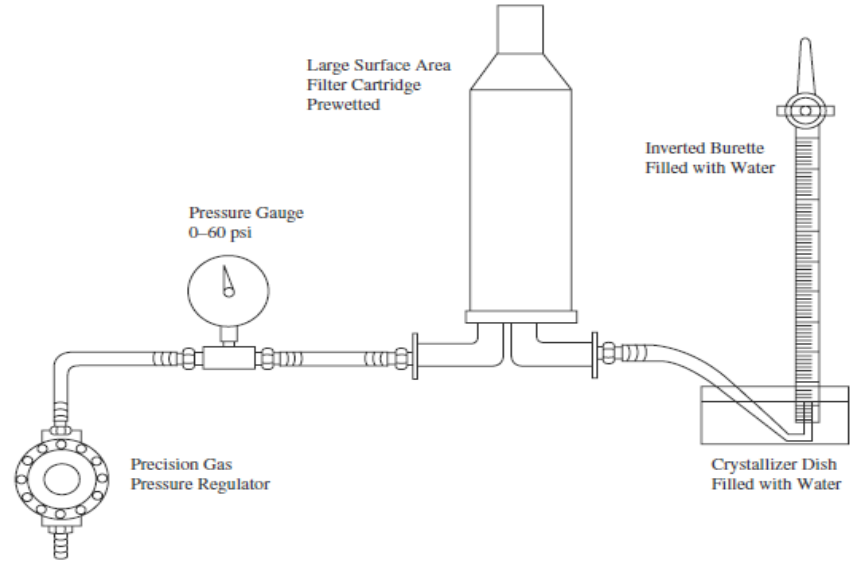
# Membrane filters

## Diffusion test / diffusive flow test

At a pressure approximately 80% of the minimum bubble point, the gas which diffuses through the membrane is measured to determine a **filter's integrity**.

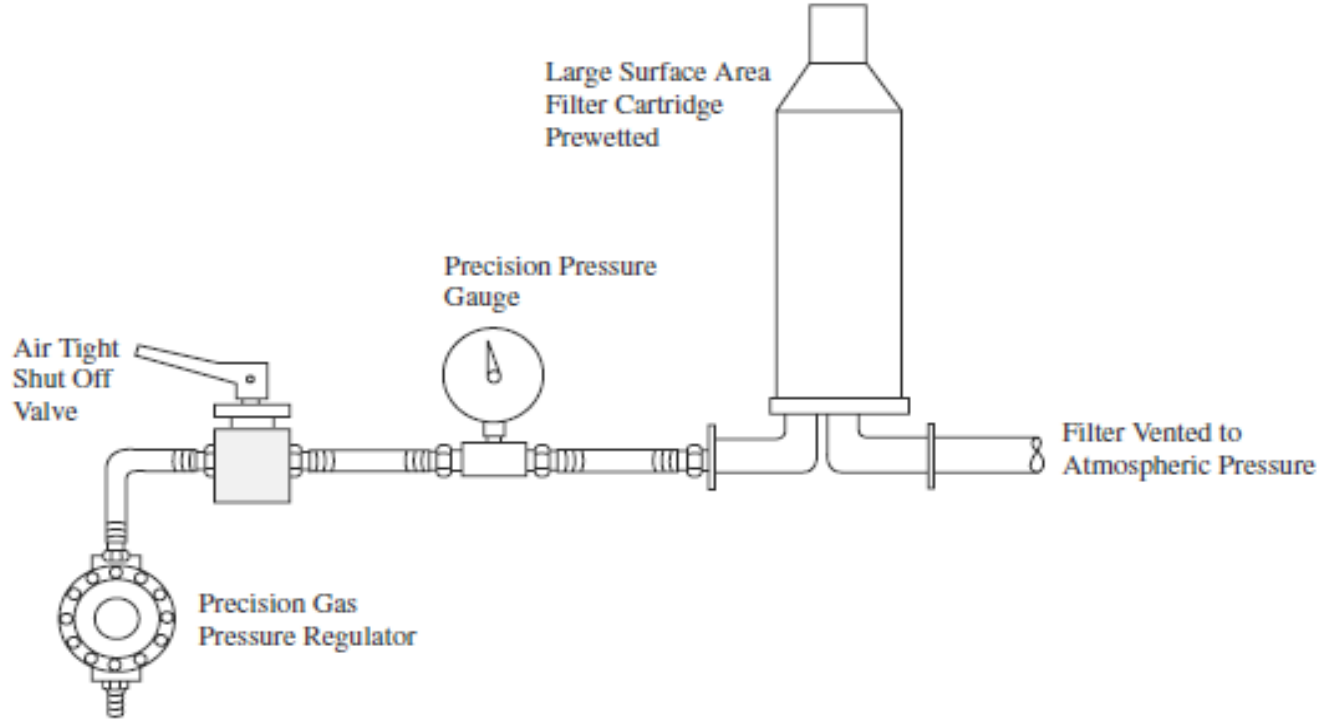
The flow of gas is very low in small area filters, but it is significant in large area filters.

Maximum diffusional flow specifications have been determined for specific membranes and devices and are used to predict bacterial retention test results.



# Membrane filters

## Pressure Hold Test



# Filtration

## application

**Solutions, syrups:** paper, textiles, glass filter

**Eye drops:** G5 sintered glass filter 1,0- 1,5  $\mu\text{m}$   $\rightarrow$  suspended contaminants

Membrane filters (elimination of microorganisms)

0,20  $\mu\text{m}$   $\rightarrow$  water based solution

0,45  $\mu\text{m}$   $\rightarrow$  oil (viscous) based solution

**Injections, infusion:** prefiltering  $\rightarrow$  suspended contaminants

~ hard porcelain, single- and multi-layer filters

endfiltering  $\rightarrow$  glass, single- and multi-layer filters, membrane filters

i.e. G5 sintered glass filter

**De-microbial filters !!!**

"those preparations, where terminal sterilization is not possible....

... 0,20  $\mu\text{m}$  membranefilter (see OGYÉI homepage)

# Reverse osmosis preparation of „aqua purificata”

The water is separated from the dissolved substances (forced through a semipermeable membrane).

The larger molecules cannot penetrate through the membrane pores (metal complexes, organic molecules).

# Reverse osmosis preparation of „aqua purificata”

The membrane material is usually **polyamide** or **cellulose-acetate**.

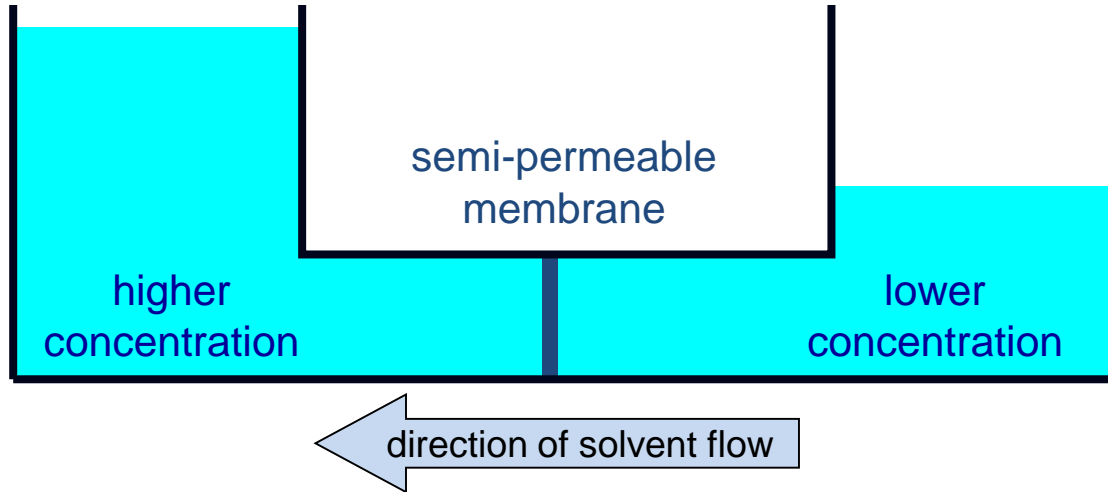
The polyamides can be applied in a large pH-range (2-11).

Carbon and metallic filter's pore size is larger, 10-100  $\mu\text{m}$ .



# Osmosis

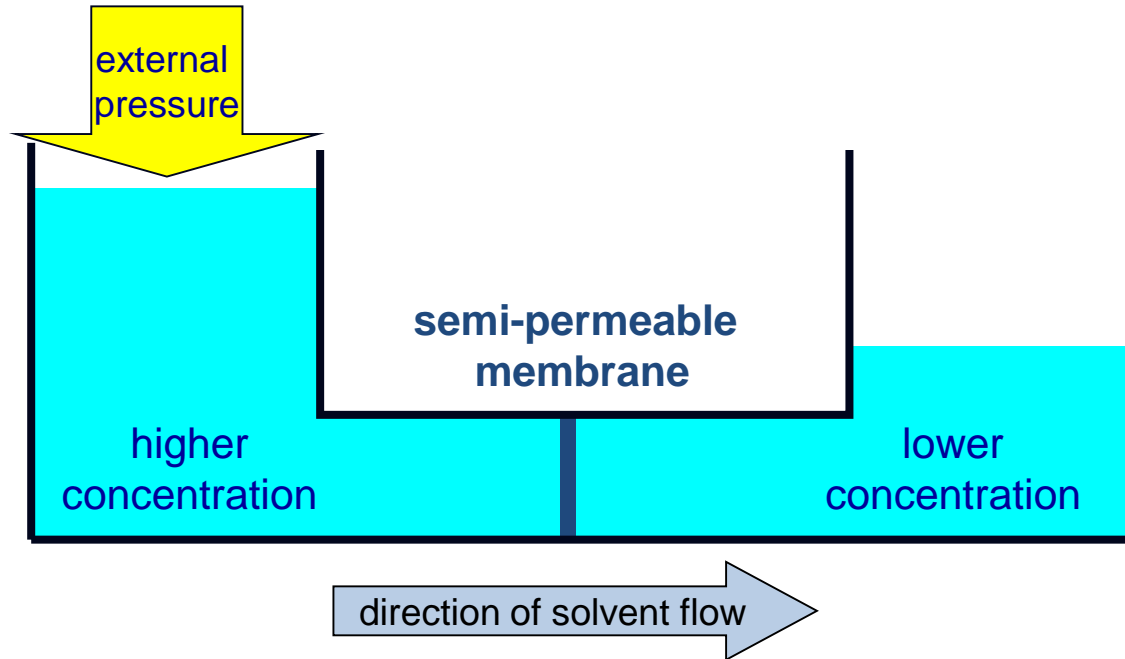
**Osmosis** is the movement of solvent molecules through a selectively permeable membrane into a region of higher solute concentration, aiming to equalize the solute concentrations on the two sides



# Osmosis

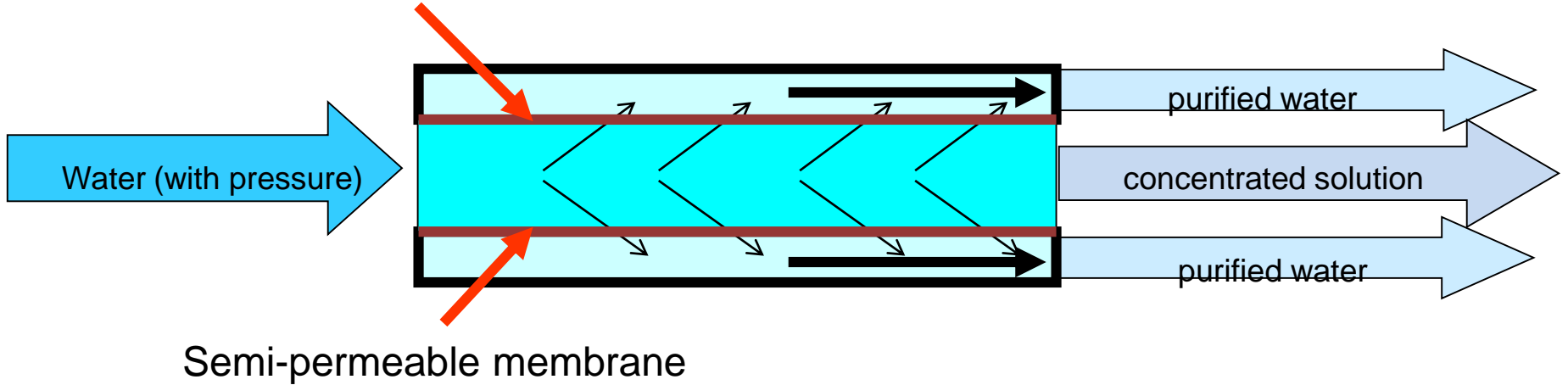
## Reverse osmosis

The direction of solvent flow can be reversed by external pressure.

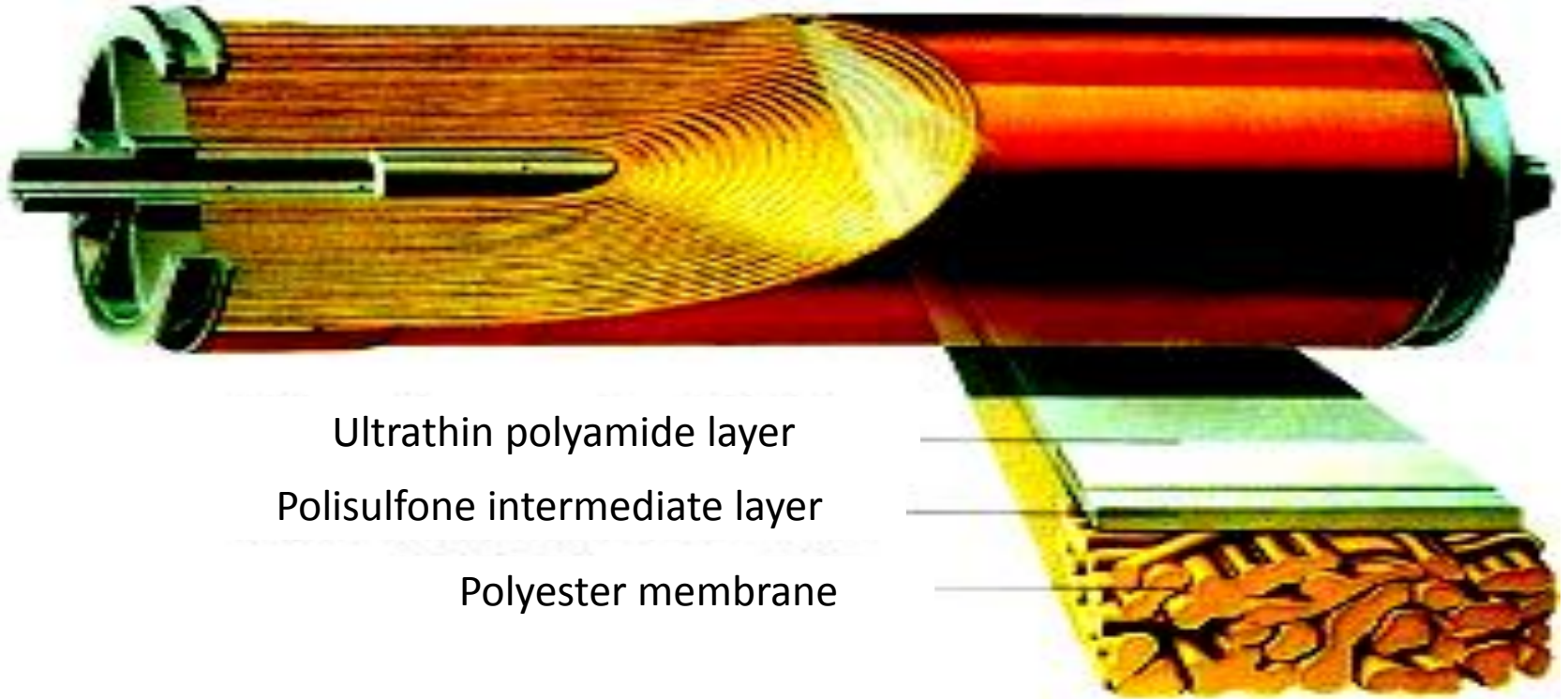


# Reverse osmosis

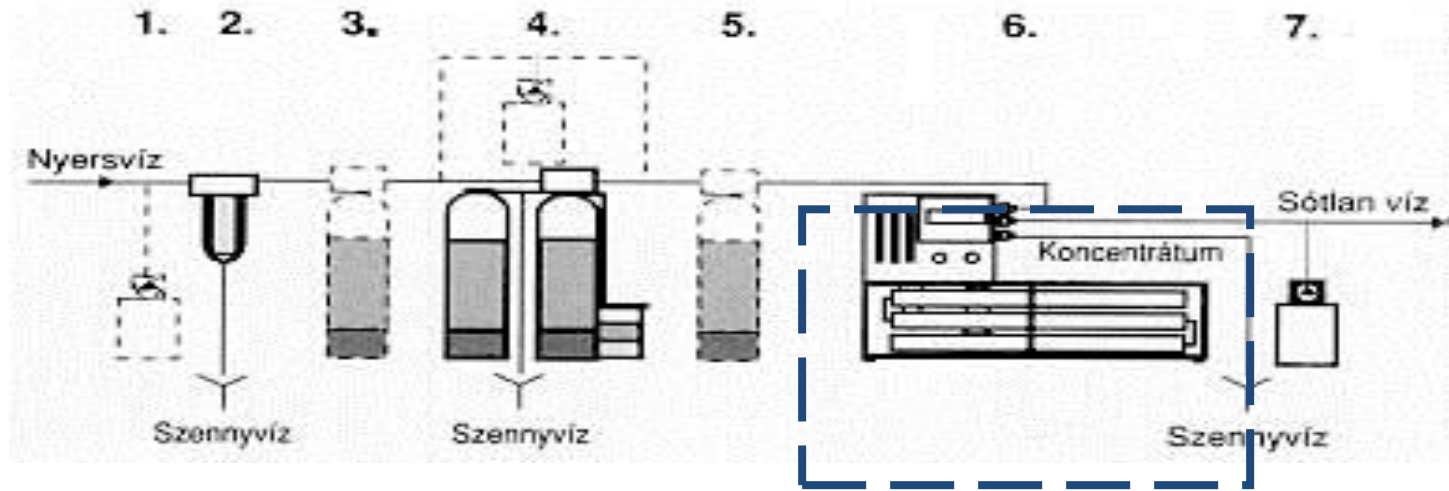
Semi-permeable membrane



# Reverse osmosis



# Reverse osmosis



1. Chemical feeder, inhibition of biological contamination
2. Suspended particles filtration, prefiltration
3. Iron removal equipment
4. Twin-column water softener – alternative: chemical feeder
5. Activated charcoal filter with high content of organic matters and free chlorine
6. *Reverse osmosis (the equipment)*
7. Chemical feeder for water conditioning

**BASIC OPERATIONS**

# **SEDIMENTATION**

# Sedimentation

Sedimentation is an **operation**,

where the phases of the disperse system (suspension) are separated

in a **density-dependent** process.

# Sedimentation

**Stokes equation** (*low concentrated suspension*)

$$v = \frac{2r^2(\zeta_1 - \zeta_2)g}{9\eta}$$

$v$  = *sedimentation speed*

$r$  = *radius of the particle*

$\zeta_1$  = *density of the dispersed phase*

$\zeta_2$  = *density of the dispersion media*

$\eta$  = *viscosity of the media*

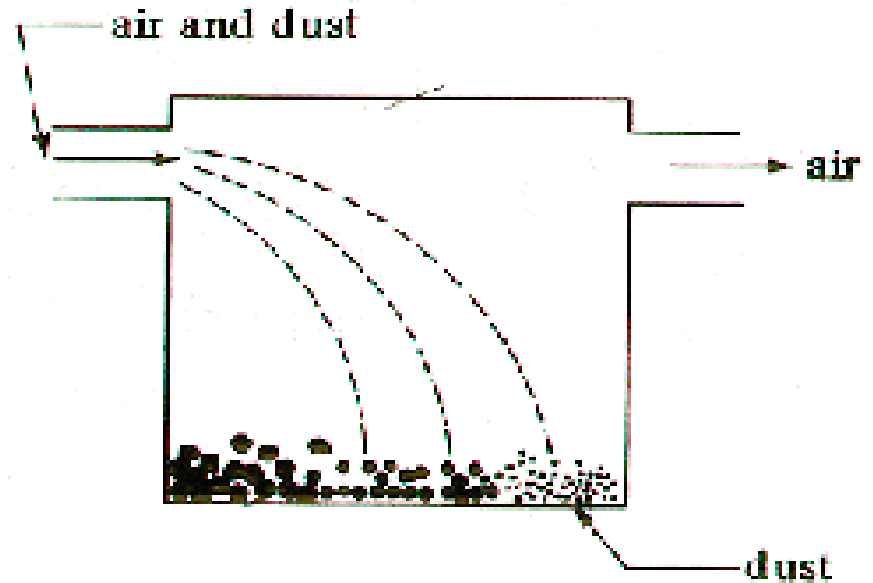
$g$  = *gravity force*



# Sedimentation

Separator container/  
dust chamber

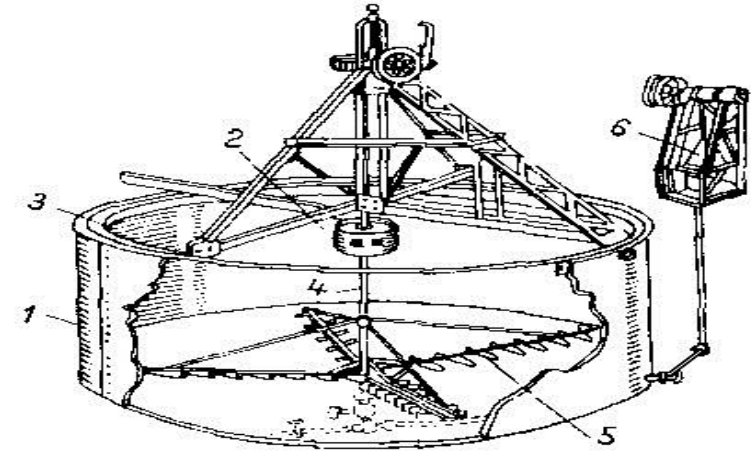
Separation of solid  
particles, powders, granules  
by air flow



# Sedimentation

## Dorr-type sedimentation box

Slow stirring, continuous removal of the sediment



- |                     |                      |
|---------------------|----------------------|
| 1. container        | 4. mixing shaft      |
| 2. feeder           | 5. mixing blades     |
| 3. drainage channel | 6. mud/sediment pump |

**BASIC OPERATIONS**

# **CENTRIFUGATION**

# Centrifugation

Centrifugation is an **operation**,

where the phases of the dispers system (suspension) are separated

in a **g-force** elevated environment.

# Centrifugation

## Application

- Separation of non-miscible liquids
- Separation of suspended solid substances from the solvent
- Remove excess fluid

# Theoretical basis of centrifugation

$$F_c = \frac{mv^2}{r}$$

$$v = r\omega$$

$$F_c = mr\omega^2$$

$v$  = *peripheral speed*

$r$  = *radius of the centrifuge's drum*

$\omega$  = *angular speed*

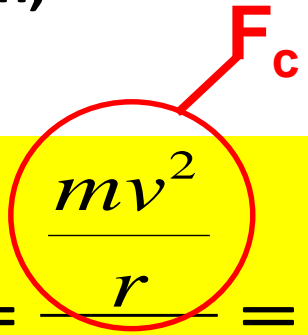
$g$  = *acceleration due to gravity*

$m$  = *mass of the particle*

$F_c$  = *centrifugal force*

# Theoretical basis of centrifugation

Separation factor of centrifugation ( $\beta$ )  
efficacy of the separation,

$$\beta = \frac{mv^2}{mg} = \frac{v^2}{rg} = \frac{r\omega^2}{g}$$


$v$  = peripheral speed

$r$  = radius of the centrifuge's drum

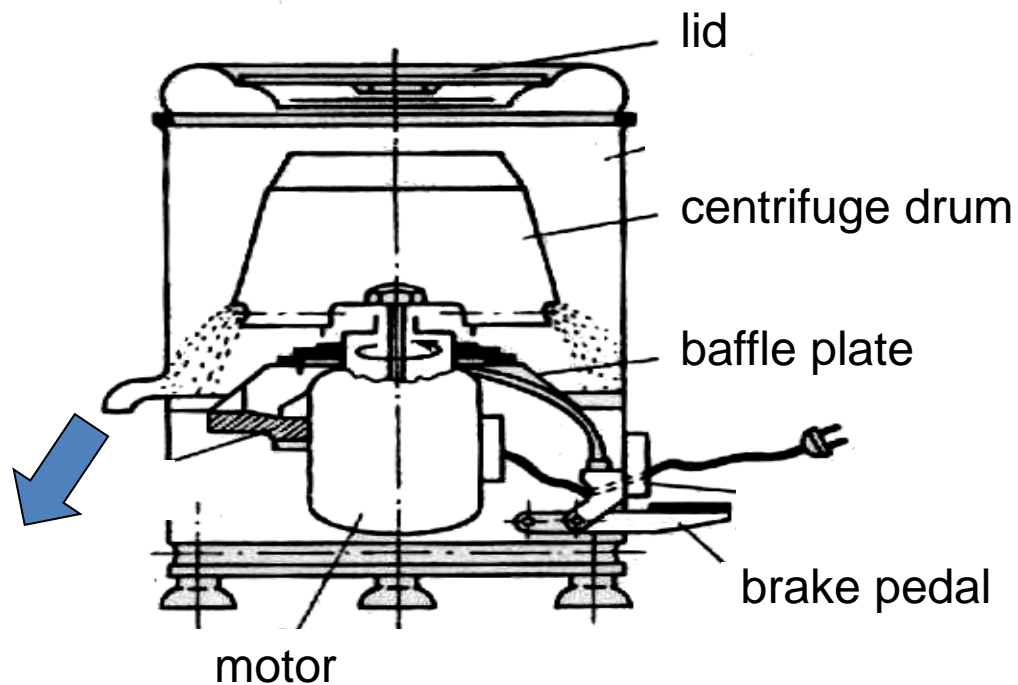
$\omega$  = angular speed

$g$  = acceleration due to gravity

$m$  = mass of the particle

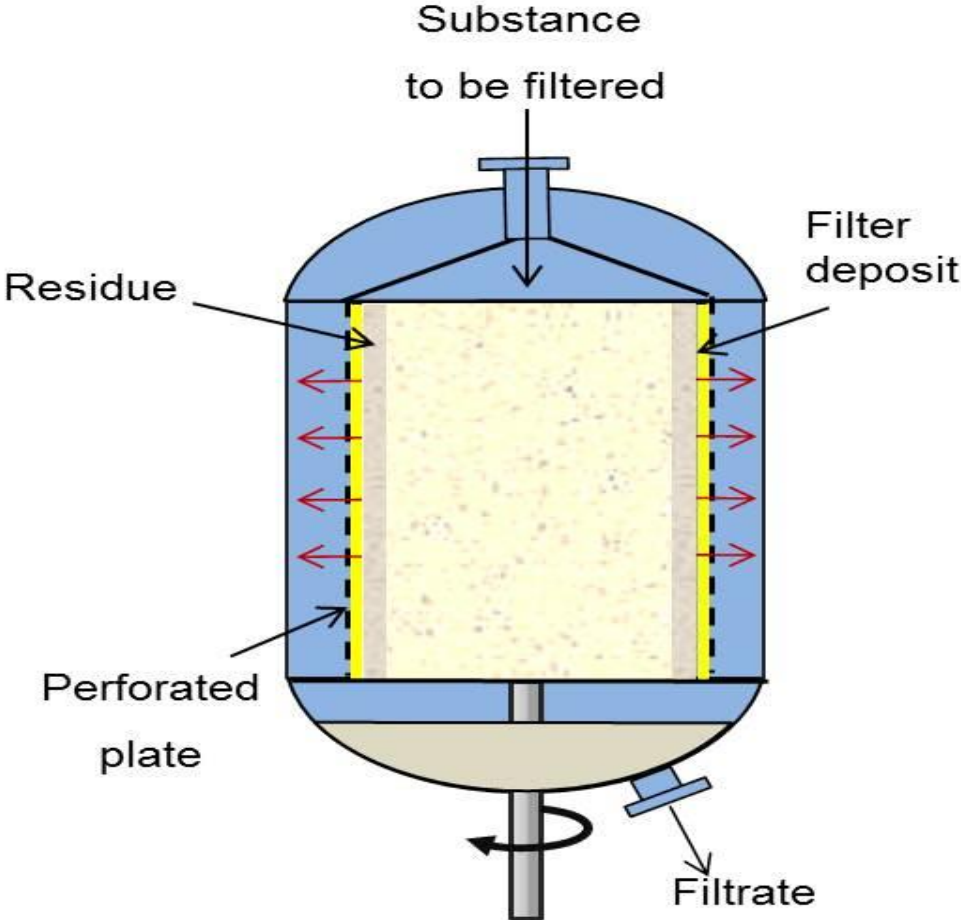
$F_c$  = centrifugal force

# Laboratory and industrial centrifuges



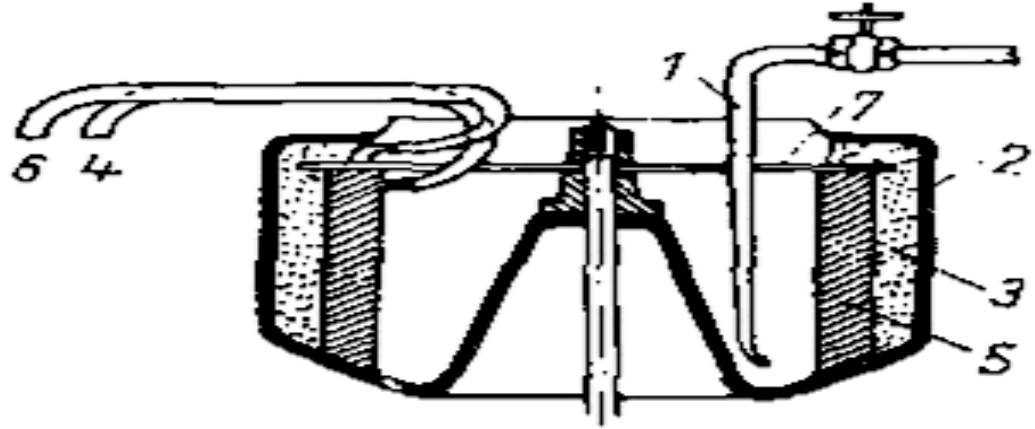


# Separator



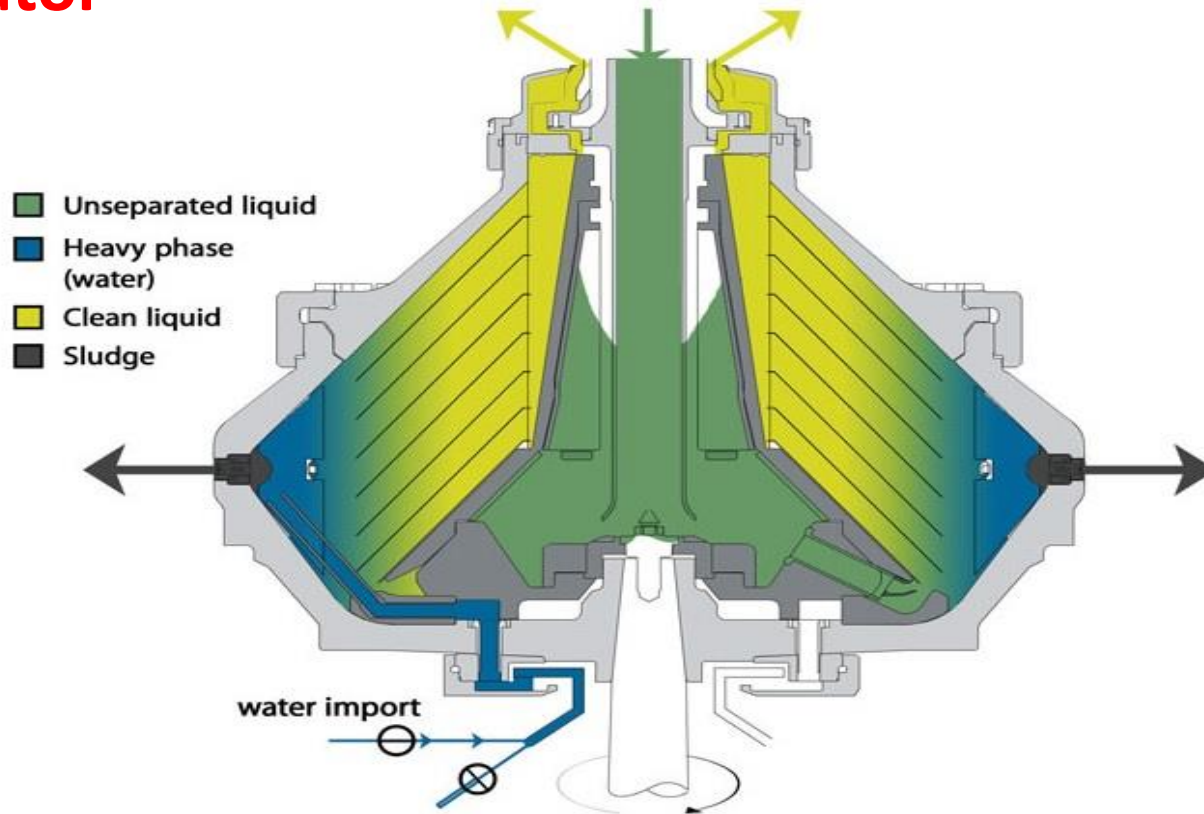
# Separator

Separation of emulsions



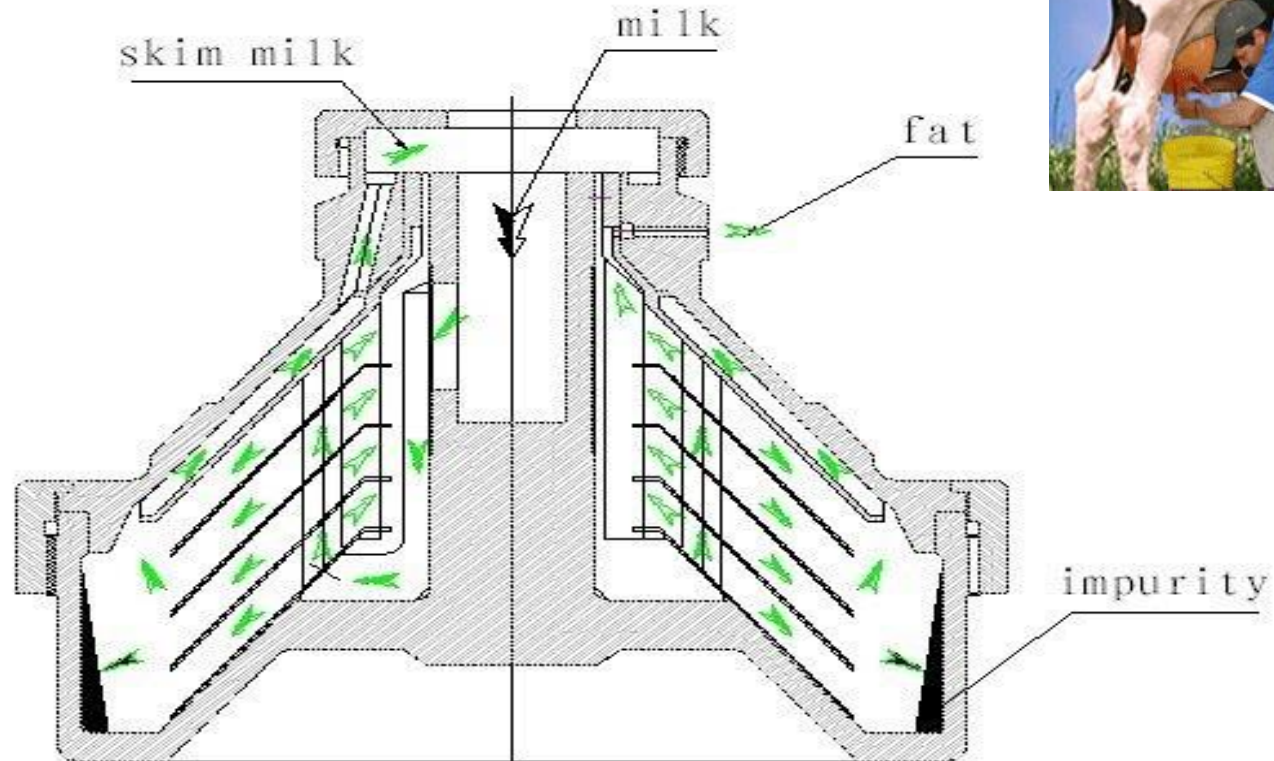
1. emulsion loading tube
2. drum
3. higher density liquid
4. lower density liquid
5. divider wall (filter)
6. drain pipe

# Disc-separator



Tapered divider walls are in the drum in which the liquid can separate in thin layer.

# Milk-separation



**The End**